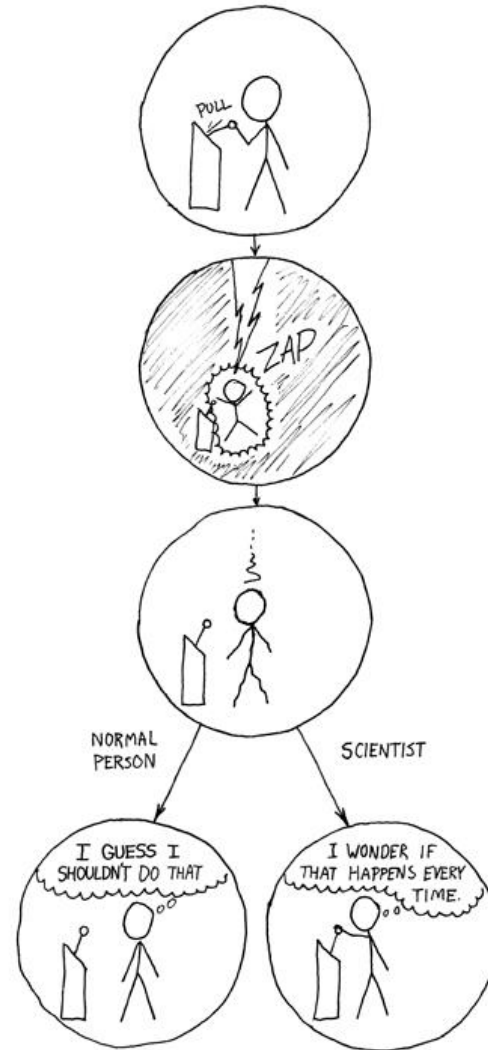


Study Design and Statistical Considerations

2019 Cape Town Targeted Quantitative Proteomics Course

Lindsay Pino

Replication



Objectives

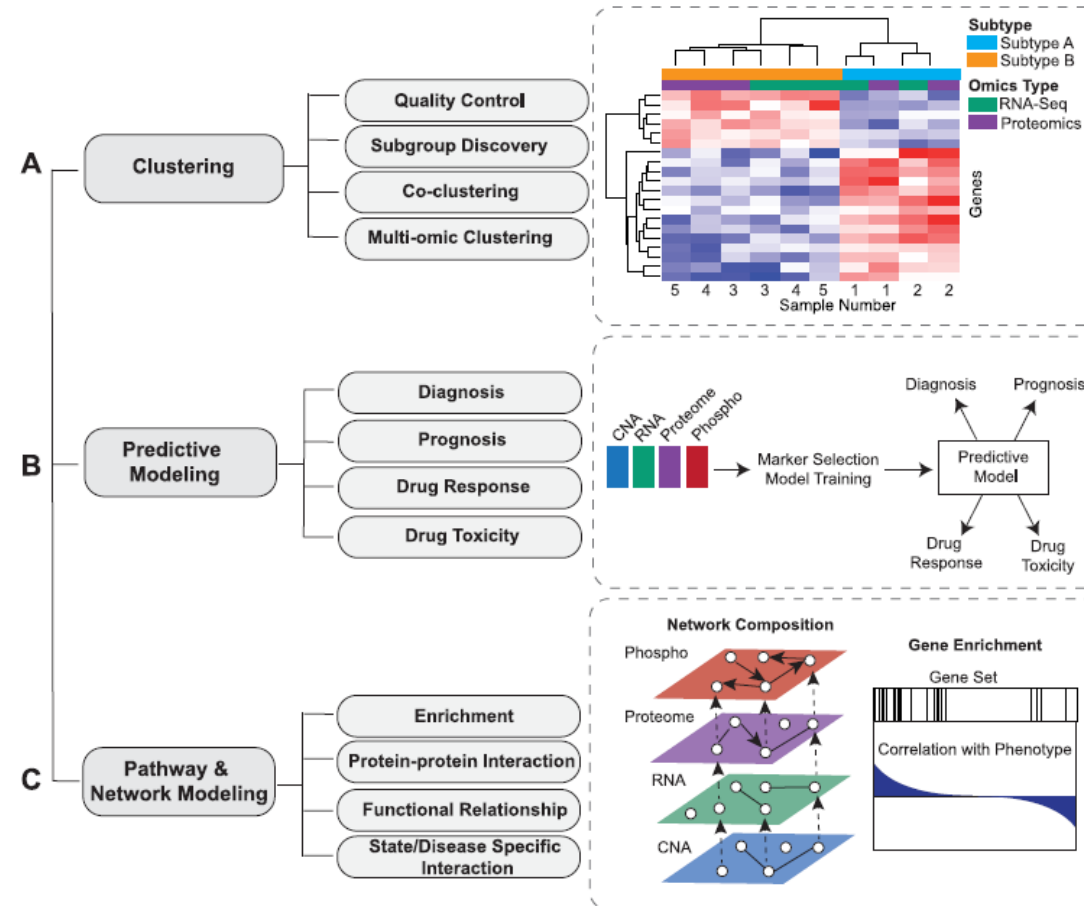
- Relate **experimental design methods** to LC-MS experiments
- Assess the **impact of variation on statistical analyses** and interpretations
- Build a **conceptual foundation** for the data processing and statistical tools contained in MSstats

Agenda

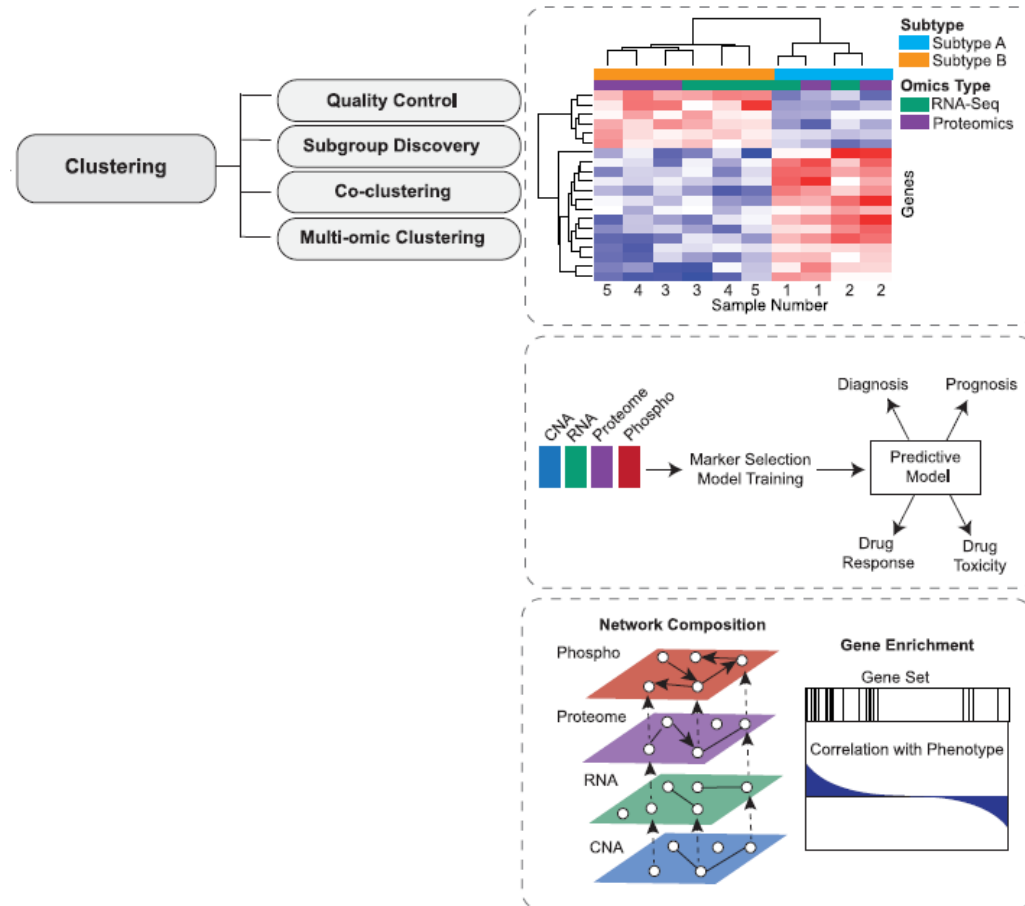
- Translating research experiments into statistical models
- Elements of good statistical/experimental design
 - Replication
 - Randomization
 - Blocking
 - Variation
- Differential abundance tests: the t-test and the ANOVA
- Statistical power: how many samples is enough and the problem with blanket fold change cut-offs

Yesterday: Group Comparisons natively within Skyline

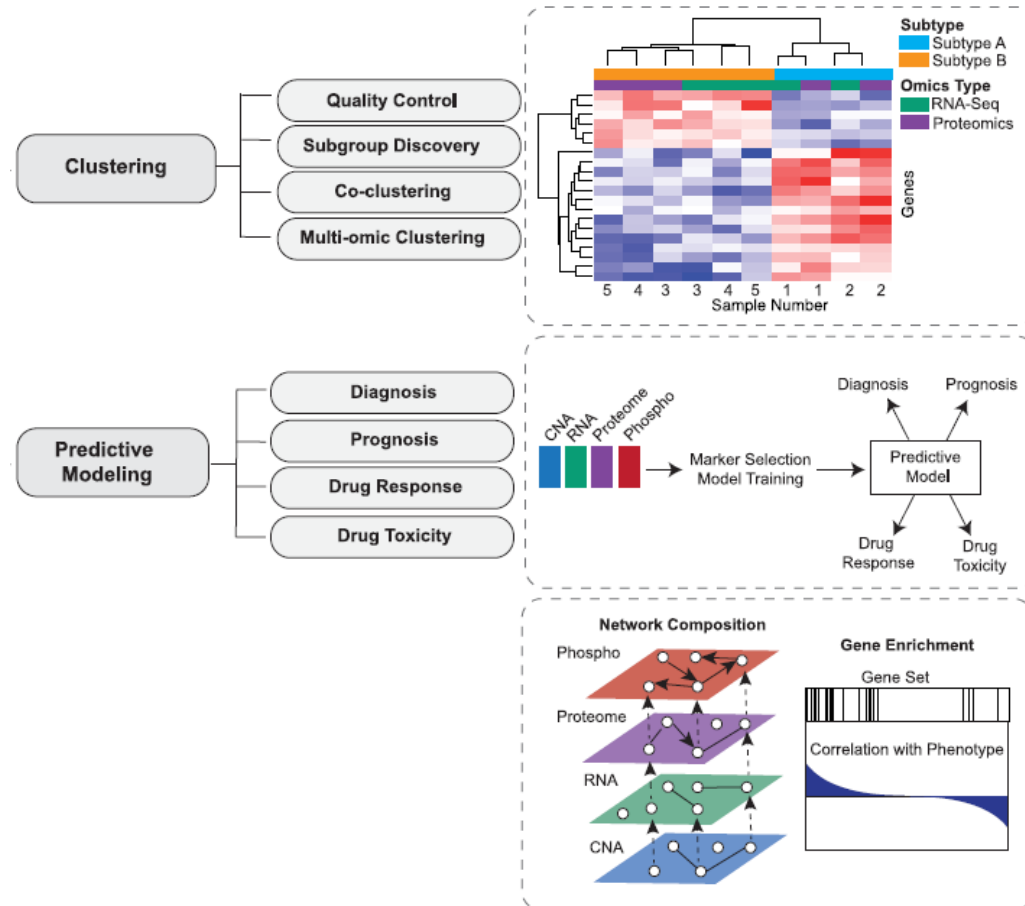
General analysis approaches for proteomics experiments



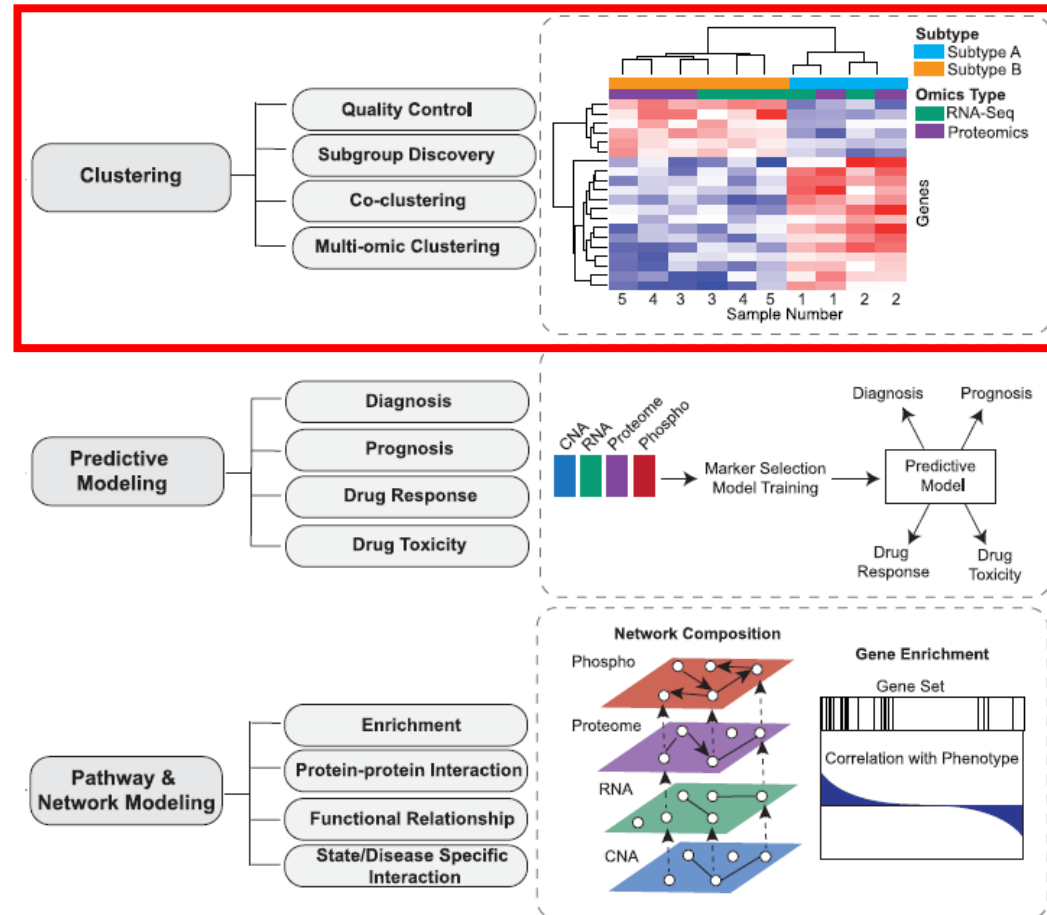
General analysis approaches for proteomics experiments



General analysis approaches for proteomics experiments



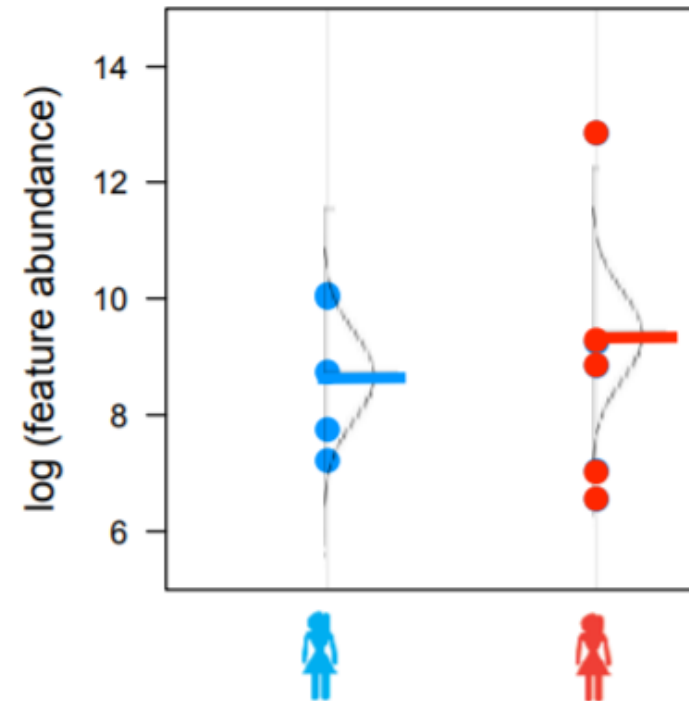
General analysis approaches for proteomics experiments



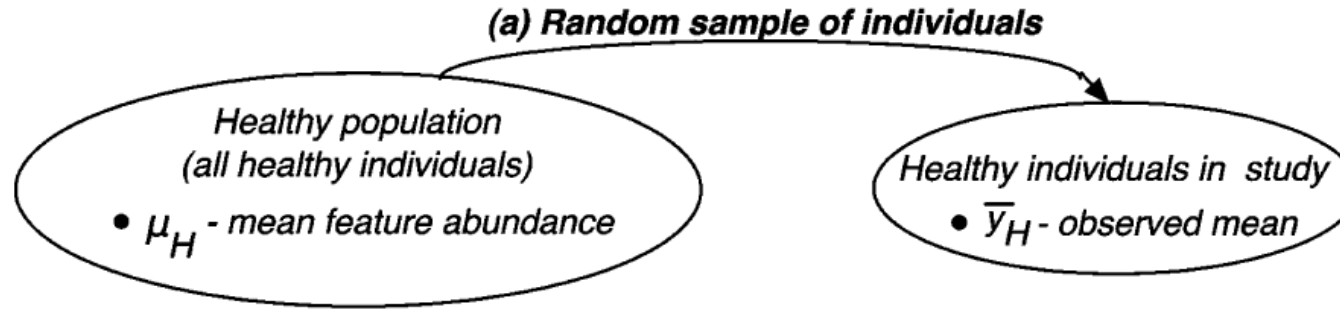
Today's focus:

Class comparison, differential analysis

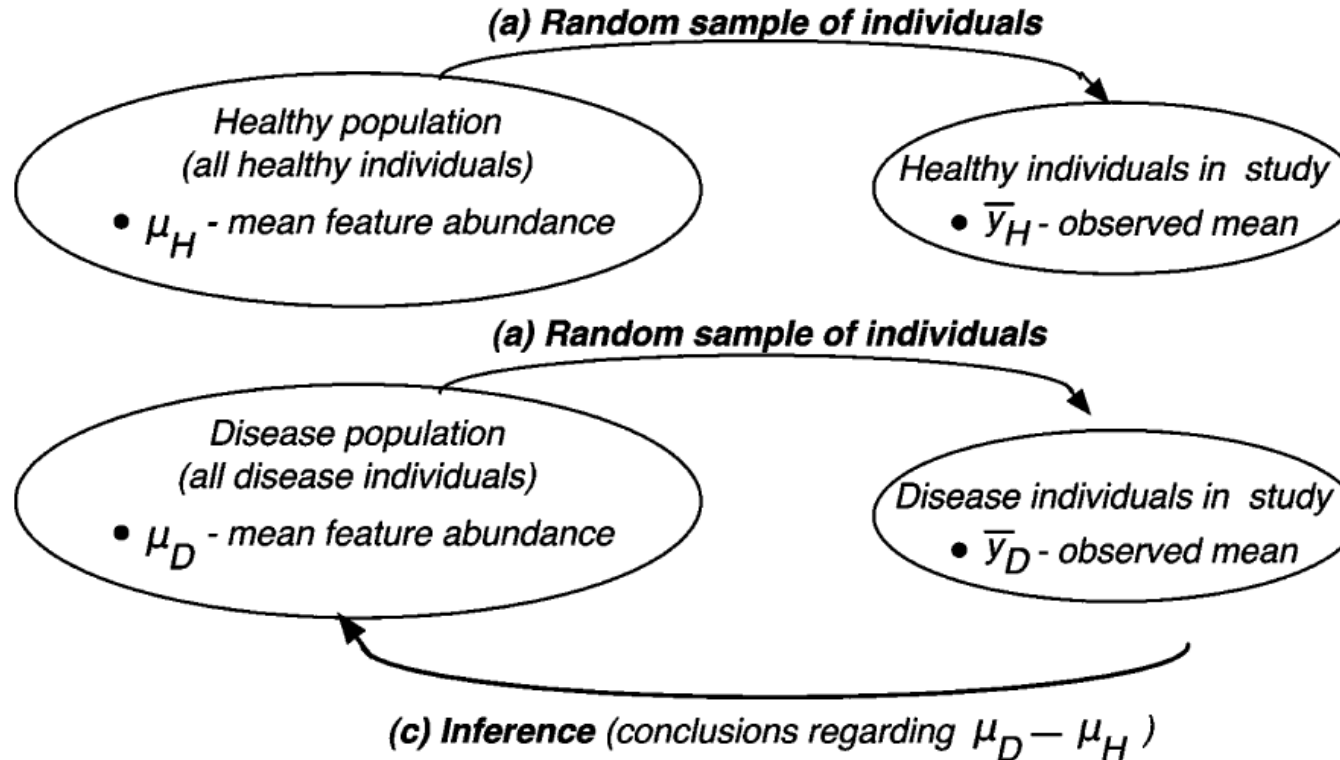
- Known class labels
 - Compare group averages
 - Report p-values, posterior probabilities etc
- Useful when compare groups of subjects
 - Best used for basic biology
 - Initial (Tier III) biomarker discovery screen



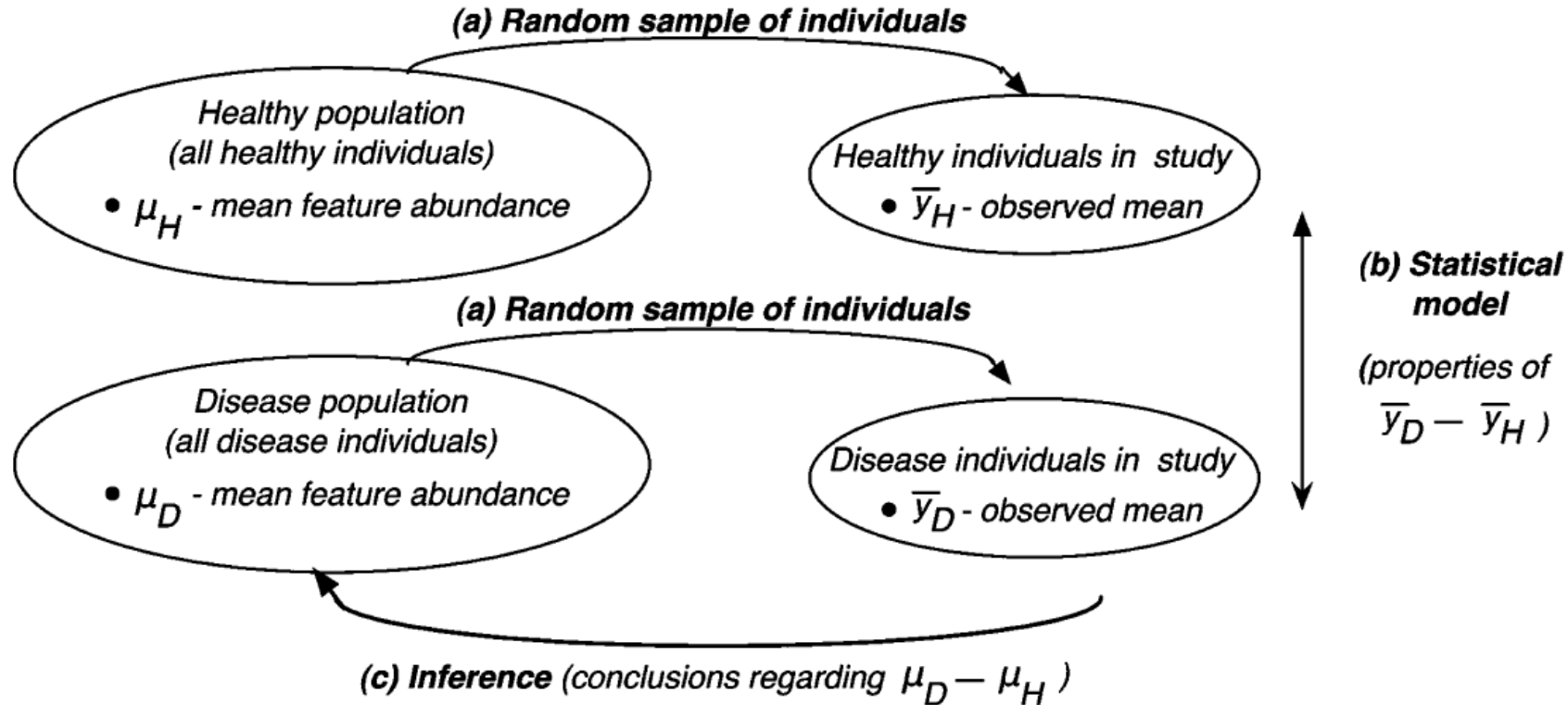
Statistical design of experiments: From data collection, to analysis, to interpretation



Statistical design of experiments: From data collection, to analysis, to interpretation



Statistical design of experiments: From data collection, to analysis, to interpretation



Agenda

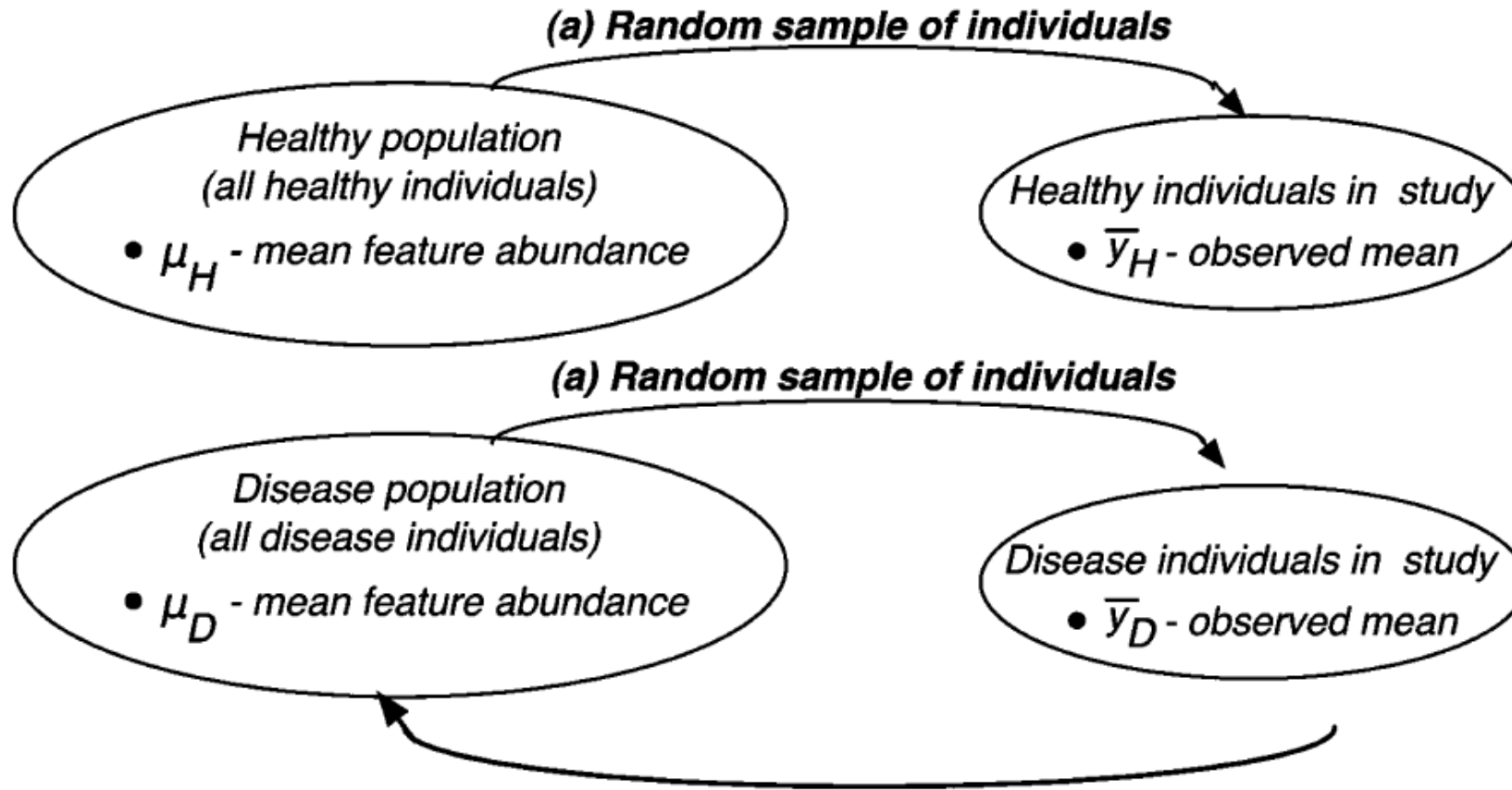
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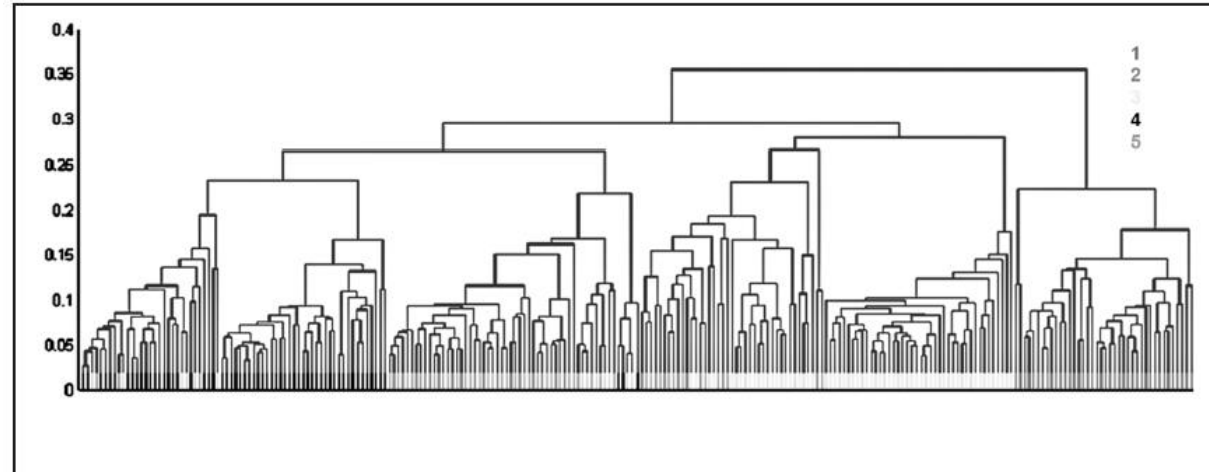
Take-Home Messages

- **Replication** is best when biological
- **Block** what you can
- **Randomize** what you can't block
- **Include all sources of variation** in models

Randomization is easy for experiments, and less so for observational studies

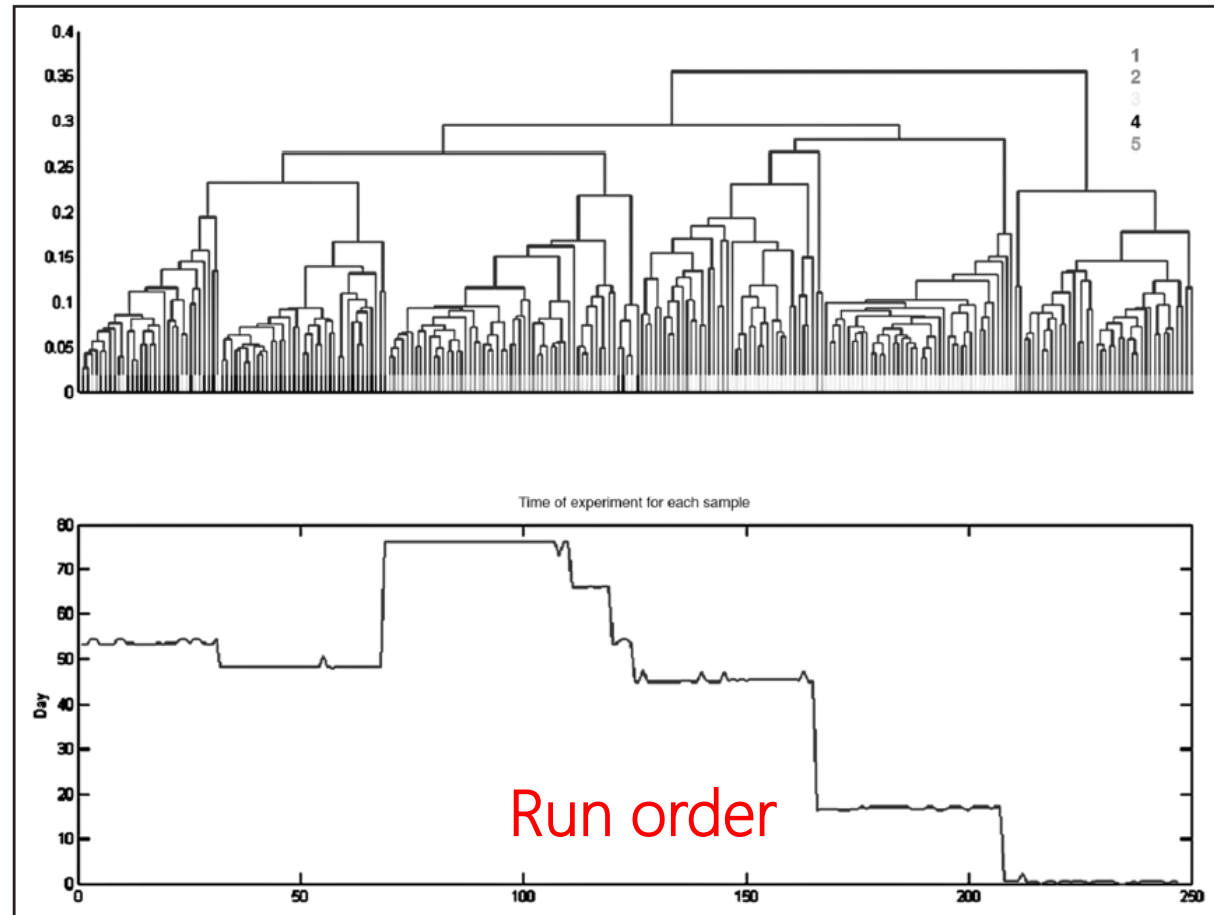


Example of randomization failure



Proteomic profiles of 5 cancer types... expected 5 groups, but analysis suggests six... what happened?

Example of randomization failure



Randomization addresses random-chance variance,
blocking helps address planned, systematic
variability

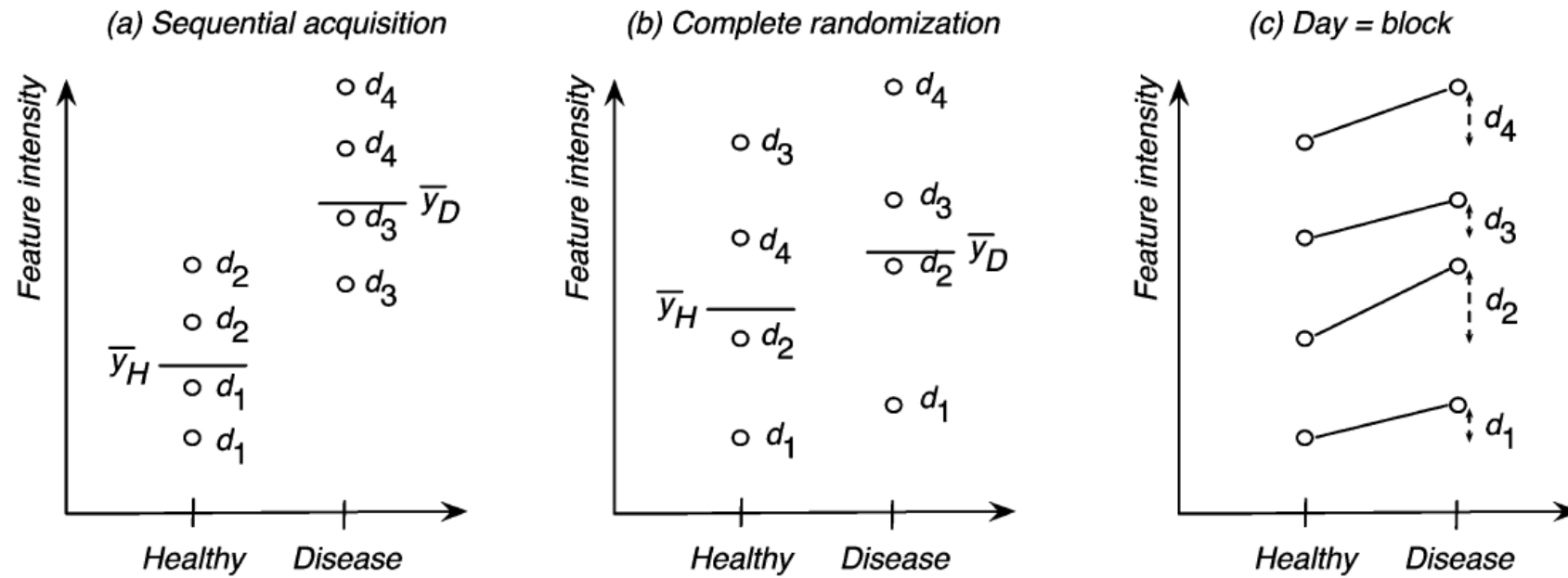
Example: blocking across time to control for variation due to instrument drift

Healthy (n=4)

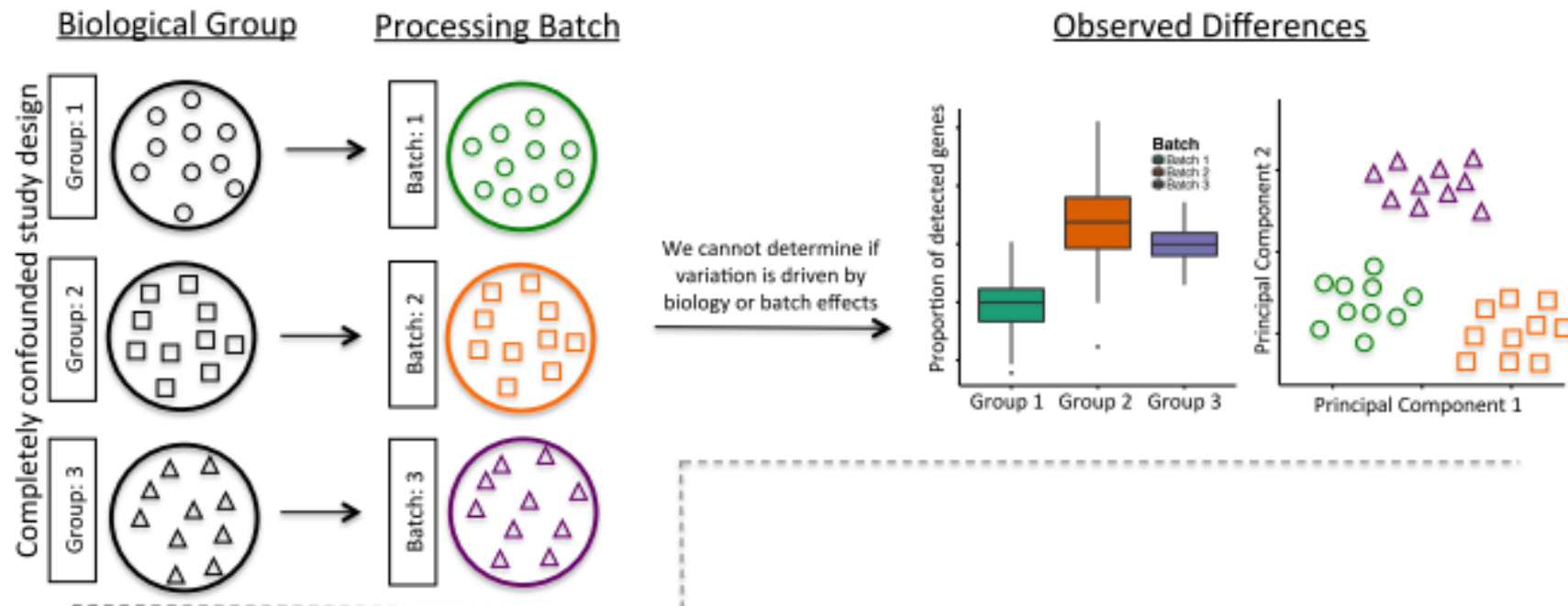
Disease (n=4)

Randomization addresses random-chance variance, **blocking** helps address planned, systematic variability

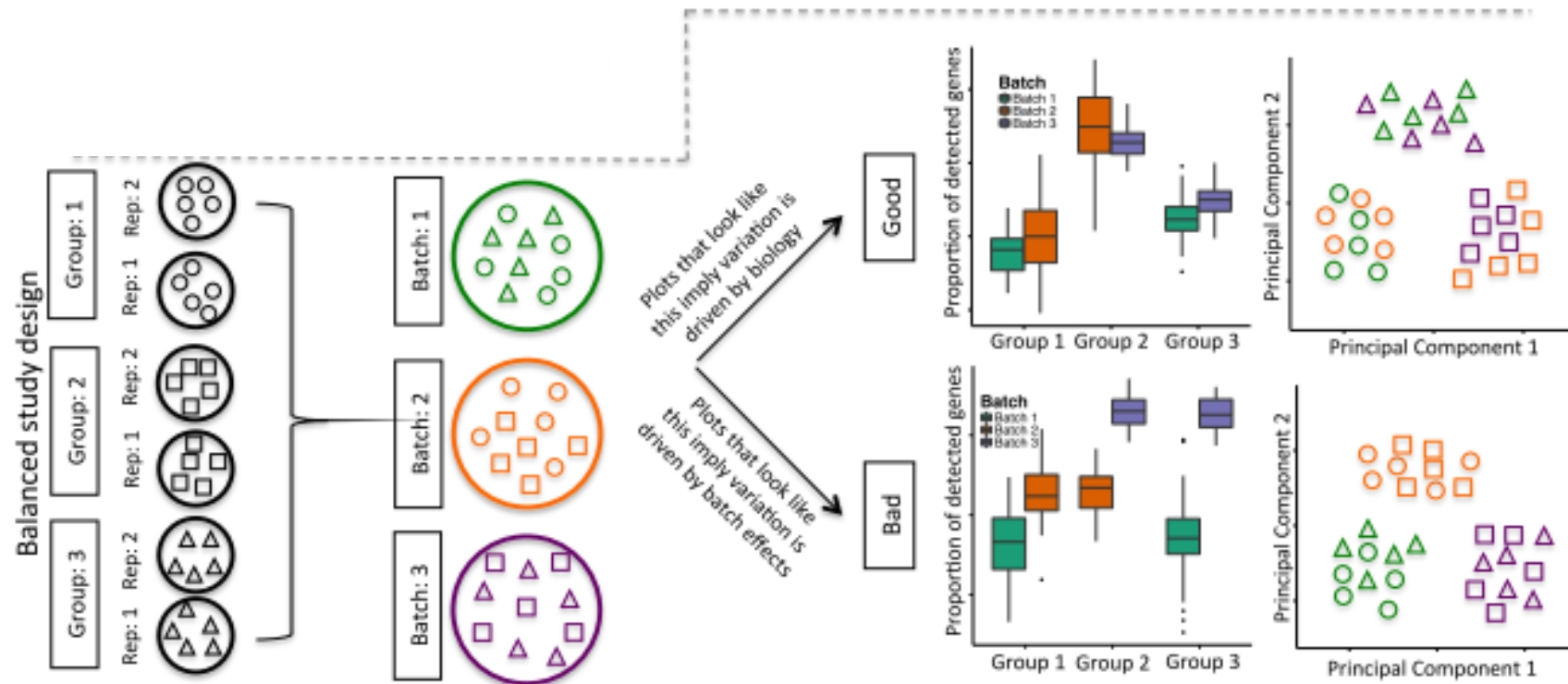
Example: blocking across time to control for variation due to instrument drift



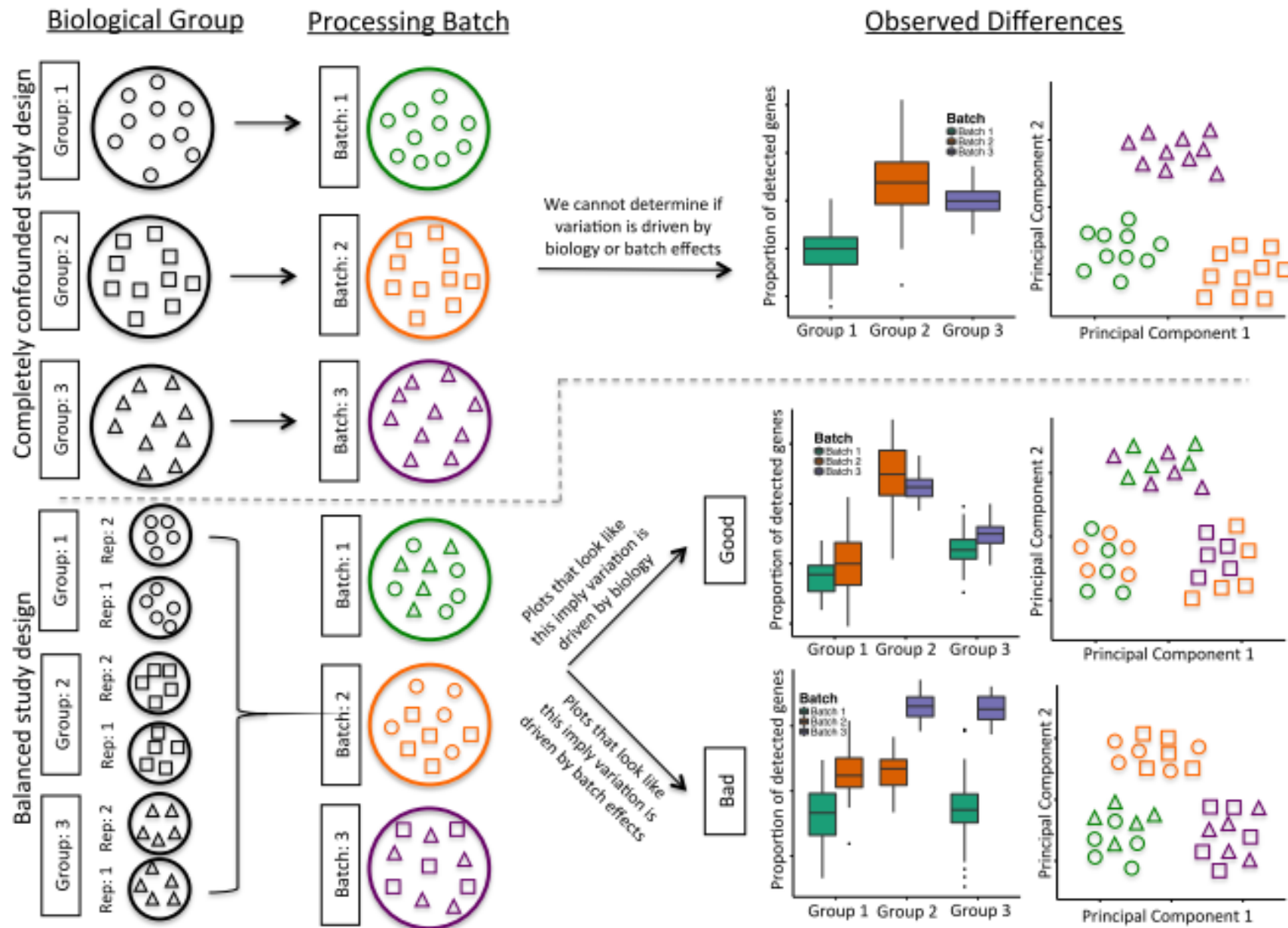
The Problem of Confounding Biological Variation and Batch Effects



The Problem of Confounding Biological Variation and Batch Effects

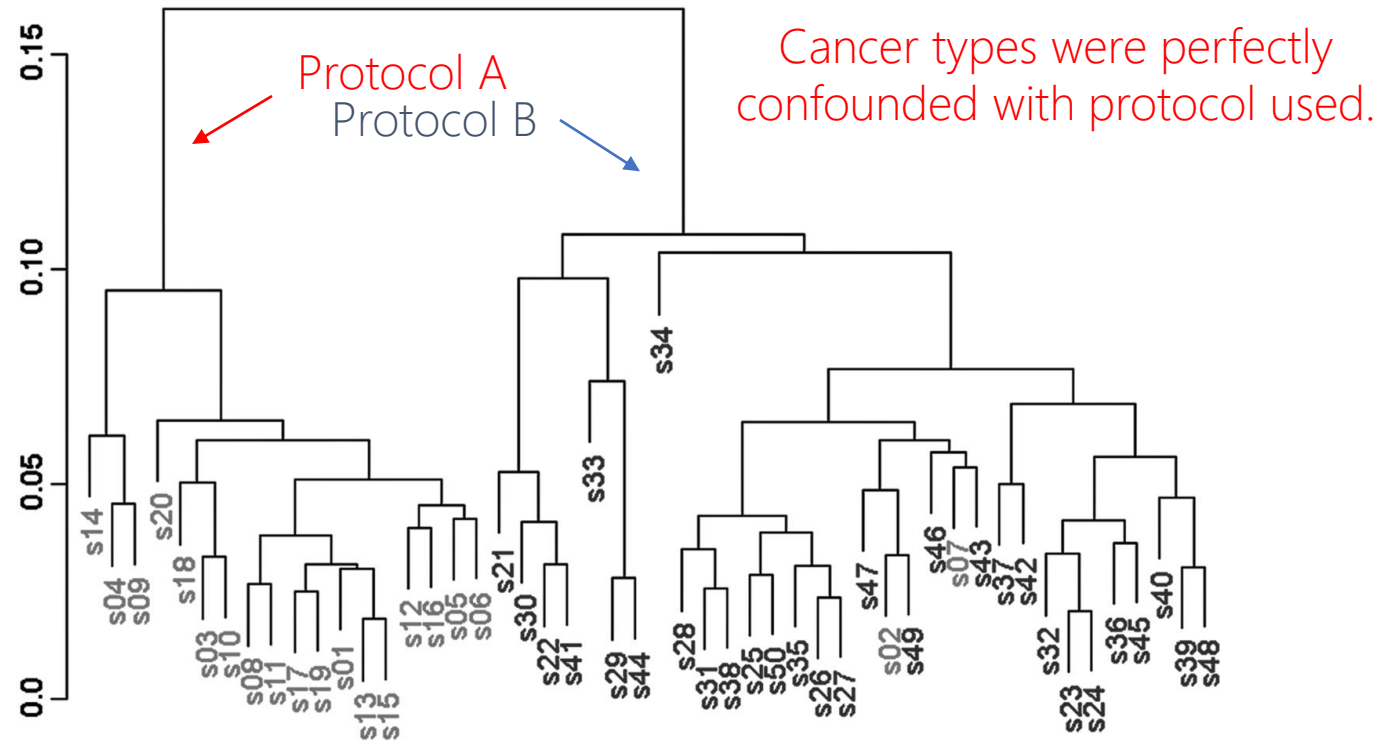


The Problem of Confounding Biological Variation and Batch Effects



Example of blocking failure

Proteomic profiles of 2 cancer types...
so this clustering checks out?



The most basic experimental design:
Completely randomized design with one replicate

Disease group	Replicate set 1
D_1	X
D_2	X
D_3	X
D_4	X

- **Advantages:** easy to set up, simple analysis, flexible
- **Disadvantages:** must be able to randomize all sources of variation (temporal or spacial)

Blocking designs that are **balanced** (all treatment comparisons are made with equal precision)

(a) Randomized Complete Block

Disease group	Replicate set 1 Block 1	Replicate set 2 Block 2	...
D_1	X	X	...
D_2	X	X	...
D_3	X	X	...
D_4	X	X	...

- **Advantages:** includes blocking to deal with heterogeneity between experimental units, straightforward statistical analysis
- **Disadvantages:** limited by block size (number of experimental units in each block must be the same as the number of treatments)

Blocking designs that are **balanced** (all treatment comparisons are made with equal precision)

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D_4	X	X	...

- **Advantages:** includes blocking to deal with heterogeneity between experimental units, straightforward statistical analysis
- **Disadvantages:** limited by block size (number of experimental units in each block must be the same as the number of treatments)

(b) Balanced Incomplete Block

Disease group	Replicate set 1					...
	Block 1	Block 2	Block 3	Block 4	Block 5	...
D_1	X	X	X	X		...
D_2	X	X	X		X	...
D_3	X	X		X	X	...
D_4	X		X	X	X	...
D_5		X	X	X	X	...

- **Advantages:** allows for smaller blocks than the RCB, still a balanced design
- **Disadvantages:** range of available designs is limited (treatments must all have equal replication, and each pair of treatments must occur together within a block exactly the same number of times over the whole experiment)

Example constructions for Balanced Incomplete Block Designs

(a) Balanced Incomplete Block

Disease group	Replicate set 1										...
	Block 1	Block 2	Block 3	Block 4	Block 5	Block 6	Block 7	Block 8	Block 9	Block 10	
D_1	X_{L_1}	X_{L_2}	X_{L_1}	X_{L_2}							...
D_2	X_{L_2}				X_{L_1}	X_{L_2}	X_{L_1}				...
D_3		X_{L_1}			X_{L_2}			X_{L_1}	X_{L_2}		...
D_4			X_{L_2}			X_{L_1}		X_{L_2}		X_{L_1}	...
D_5				X_{L_1}			X_{L_2}		X_{L_1}	X_{L_2}	...

Note: treatments must all have equal replication, and each pair of treatments must occur together within a block the same number of times over the whole experiment

(b) Reference

Disease group	Replicate set 1					...
	Block 1	Block 2	Block 3	Block 4	Block 5	
R	R_{L_1}	R_{L_1}	R_{L_1}	R_{L_1}	R_{L_1}	...
D_1	X_{L_2}					...
D_2		X_{L_2}				...
D_3			X_{L_2}			...
D_4				X_{L_2}		...
D_5					X_{L_2}	...

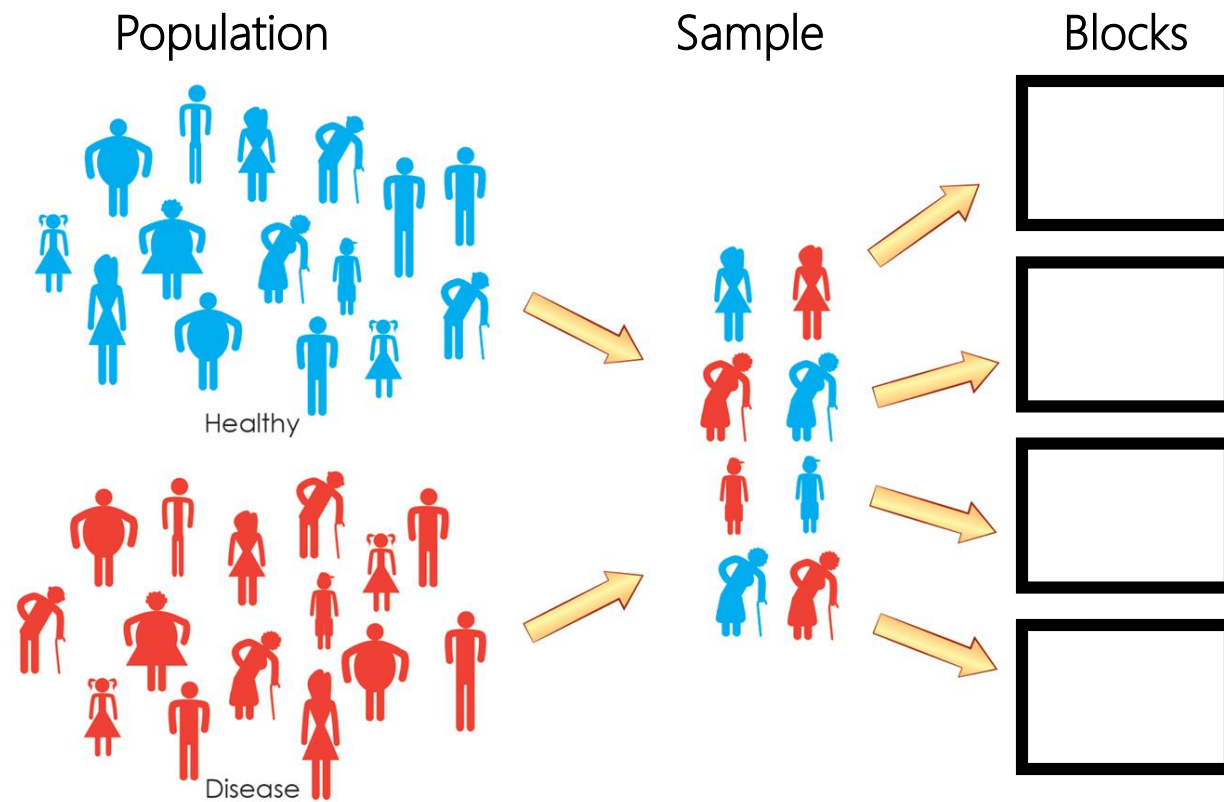
Here, the reference sample is of no scientific interest in the study, but adds experimental noise to the data to control the between-experiment variation

(c) Loop

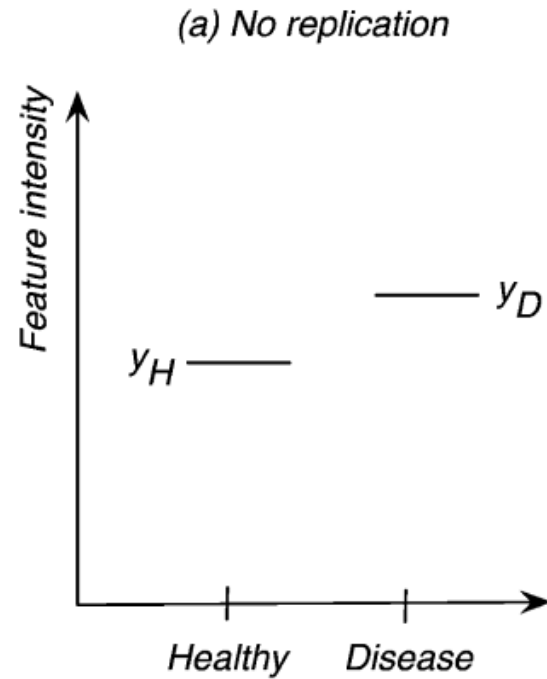
Disease group	Replicate set 1					...
	Block 1	Block 2	Block 3	Block 4	Block 5	
D_1	X_{L_1}				X_{L_2}	...
D_2	X_{L_2}	X_{L_1}				...
D_3		X_{L_2}	X_{L_1}			...
D_4			X_{L_2}	X_{L_1}		...
D_5				X_{L_2}	X_{L_1}	...

Here, the aim is to compare treatment conditions and there is no need for a control because each treatment is compared directly with another

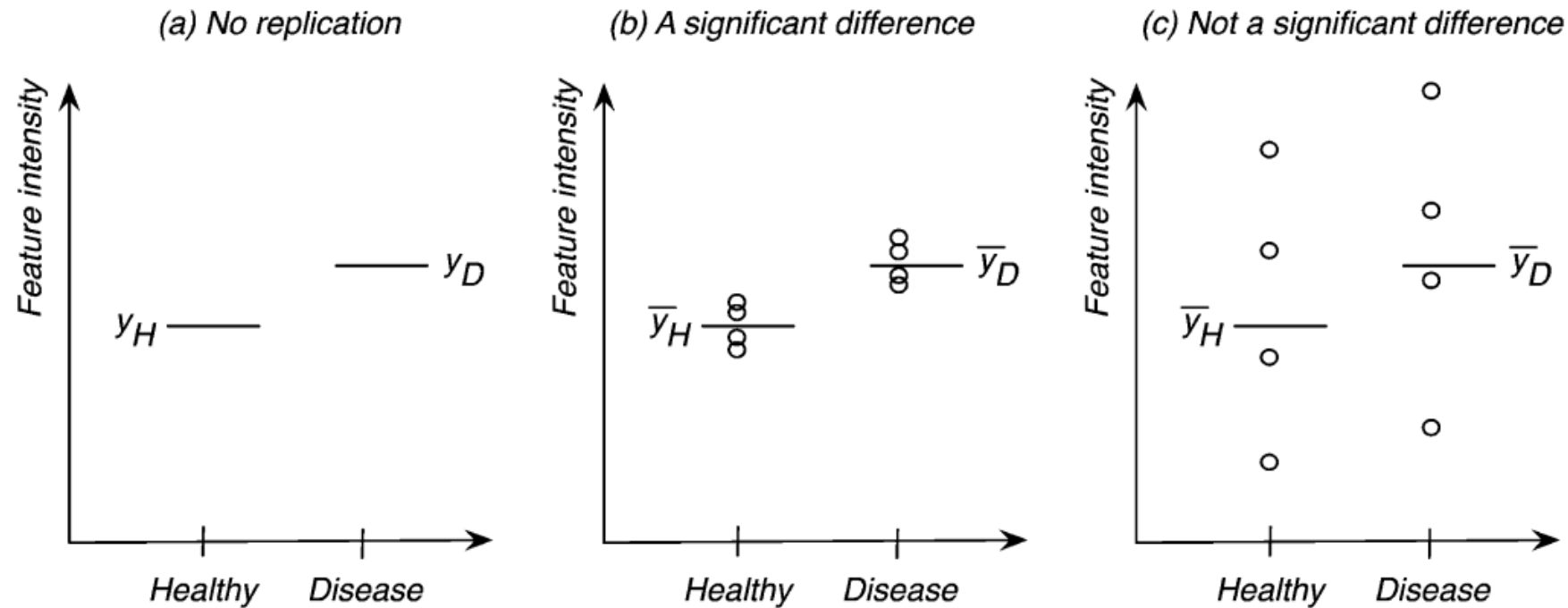
Thought exercise:
How should these eight samples be blocked?



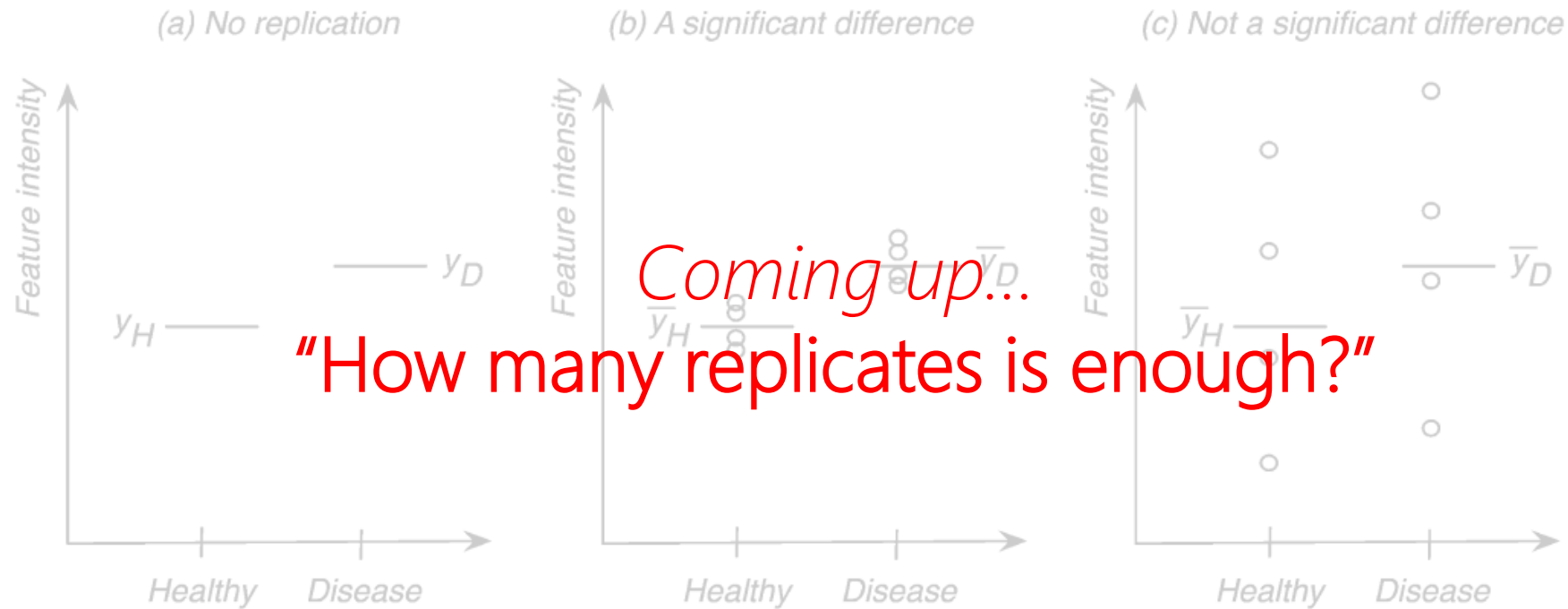
Replication assesses variation due to random chance,
ensures reliability of conclusions



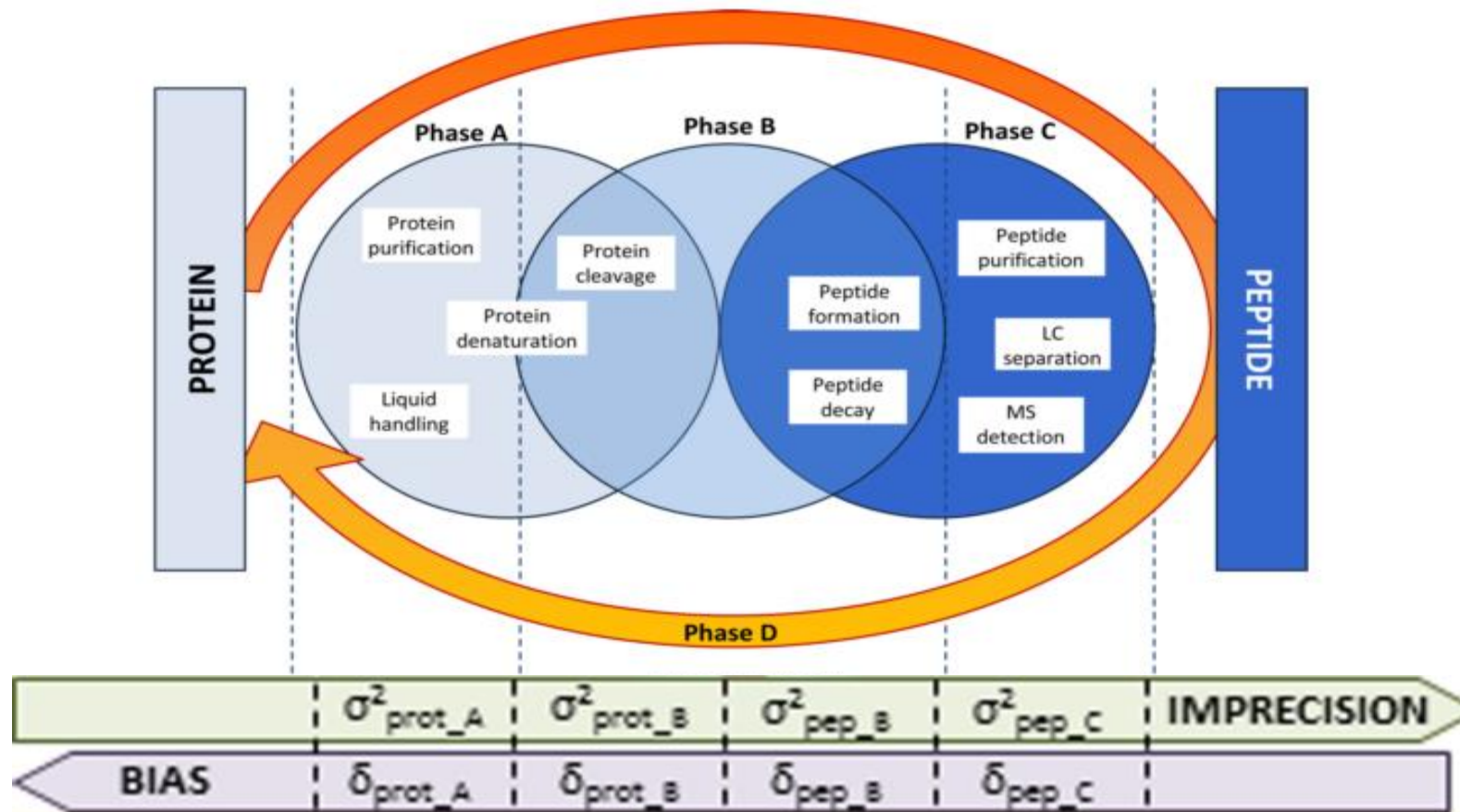
Replication assesses variation due to random chance, ensures reliability of conclusions



Replication assesses variation due to random chance, ensures reliability of conclusions



What are common sources of variation in proteomics? How can they be avoided or at least accounted for?



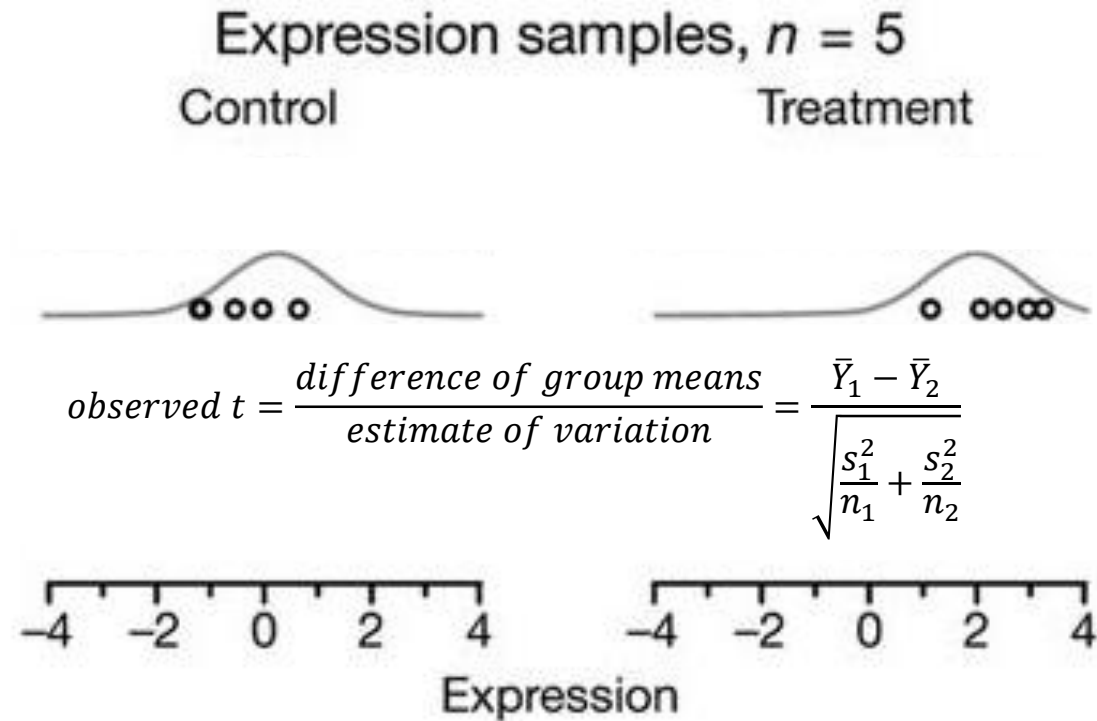
Bias: systematic errors in our conclusions

Agenda

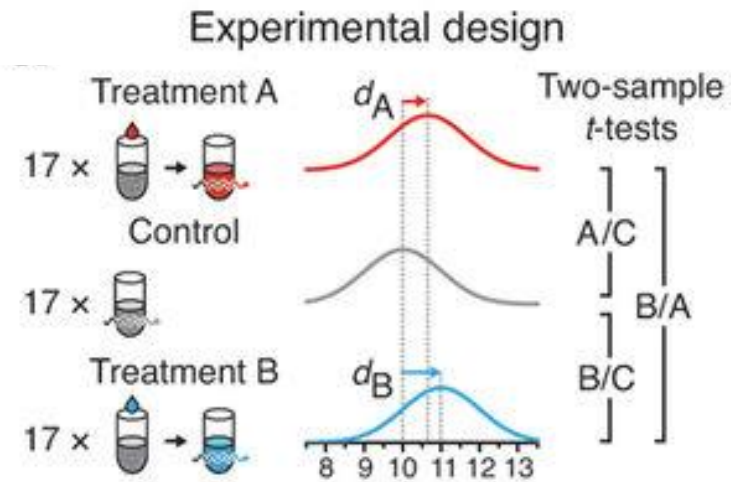
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Earlier Today: Group Comparisons natively within Skyline

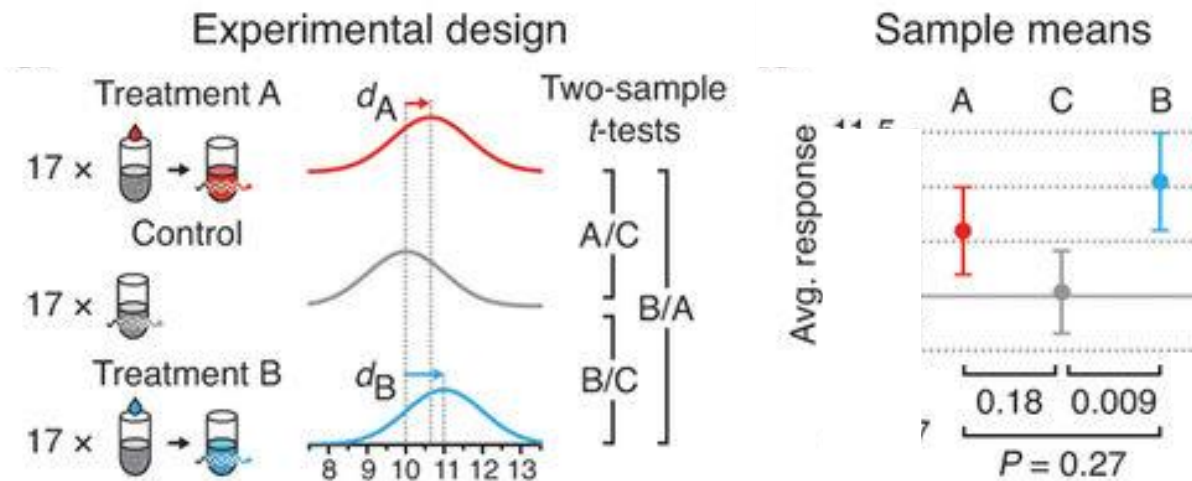
Differential abundance by the two-sample t-test



Multiple two-sample t-test for more complex experimental designs

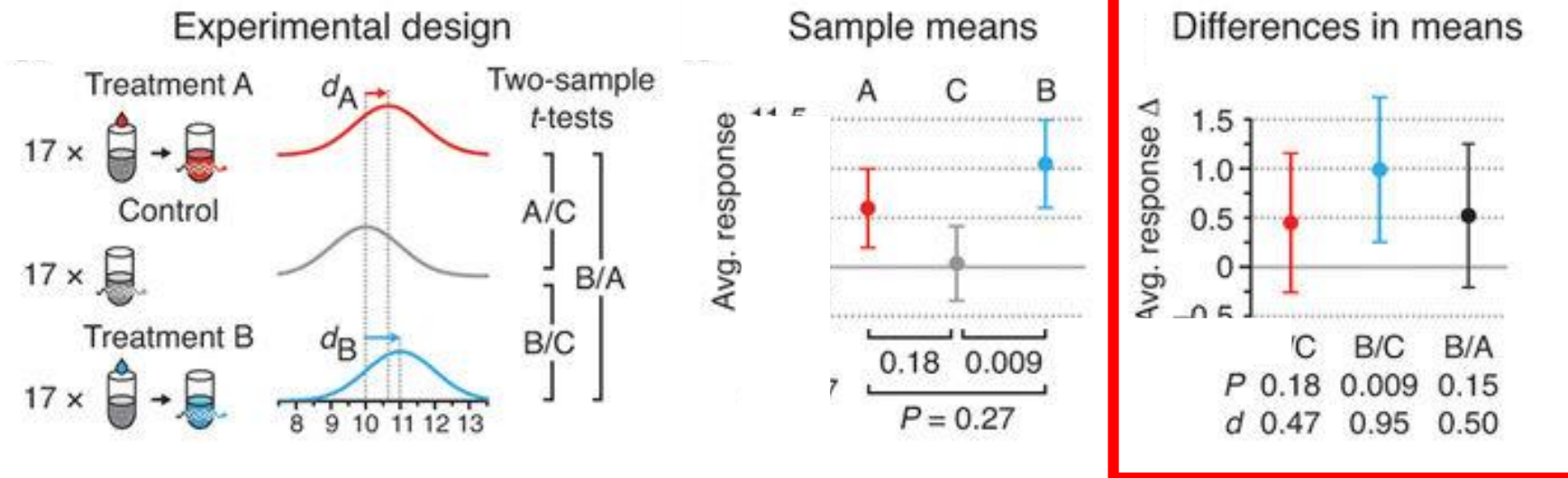


Multiple two-sample t-test for more complex experimental designs



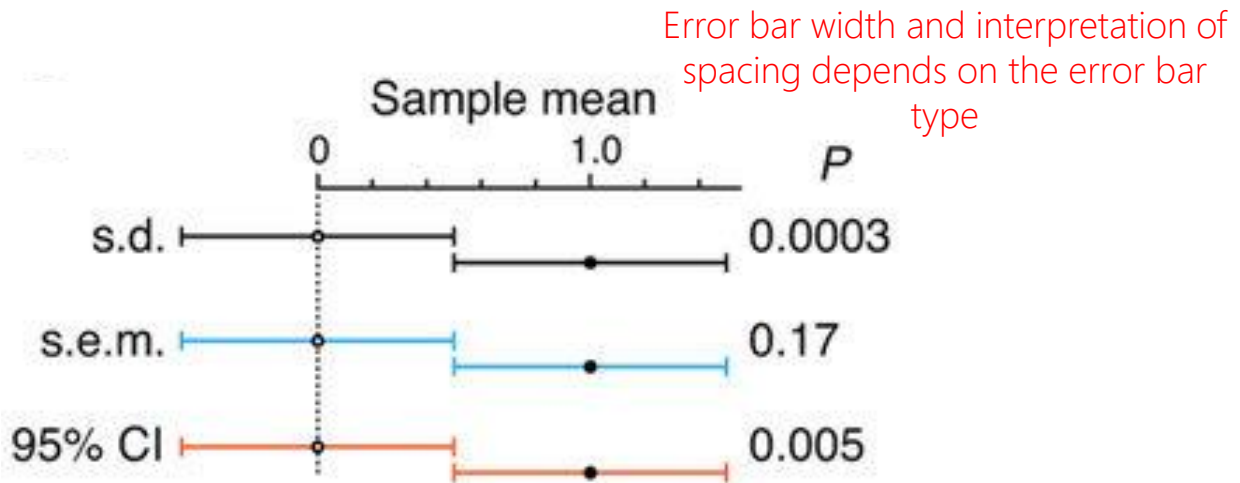
Multiple two-sample t-test for more complex experimental designs

But these error bars overlap?

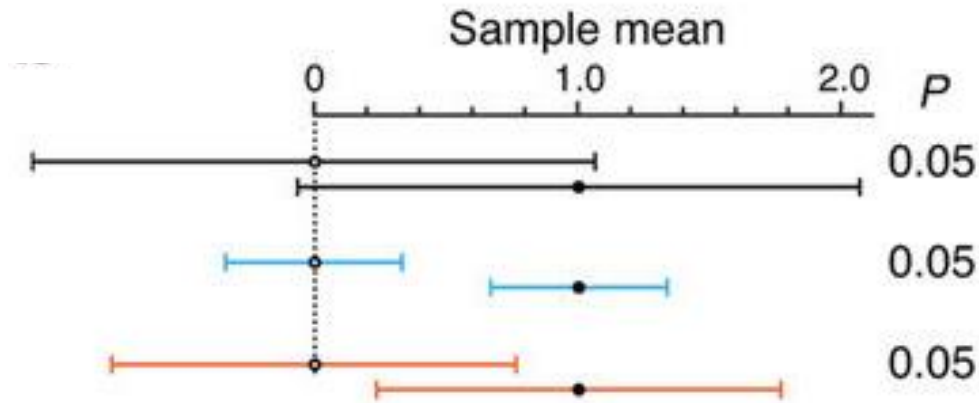


Absence (or presence) of error bar overlap does not always mean statistical significance

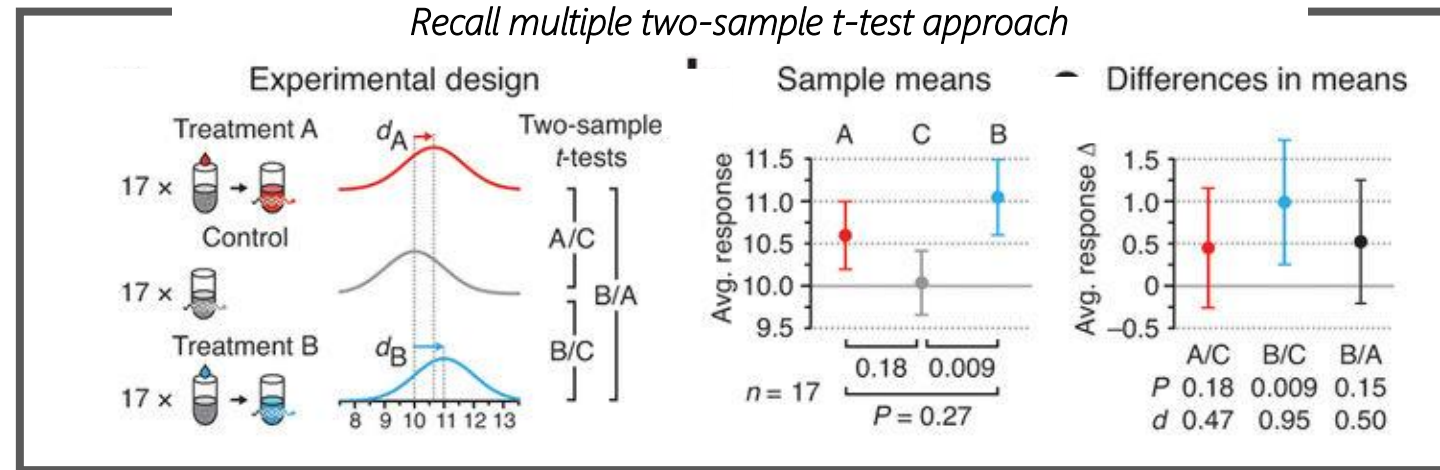
Scaled error bars,
Unequal p-values



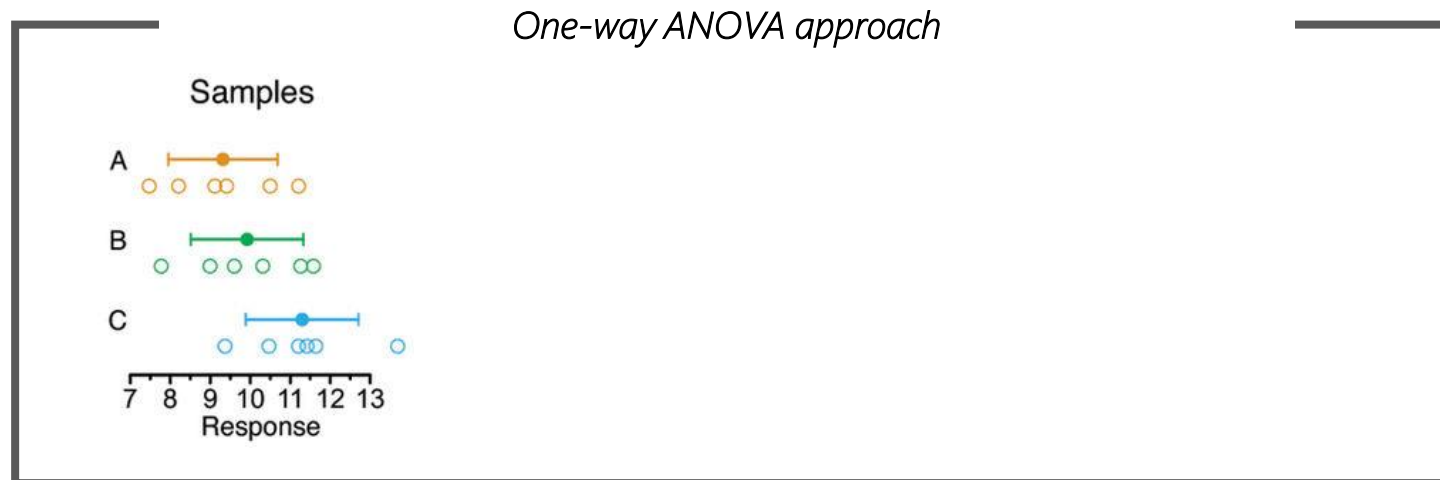
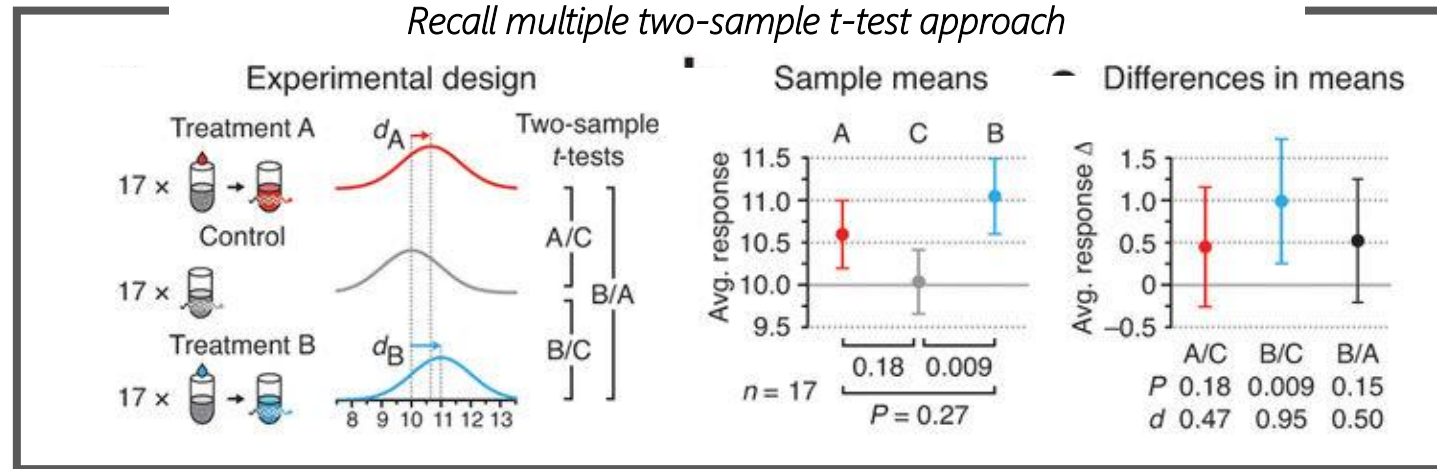
Unscaled error bars,
Equal p-values



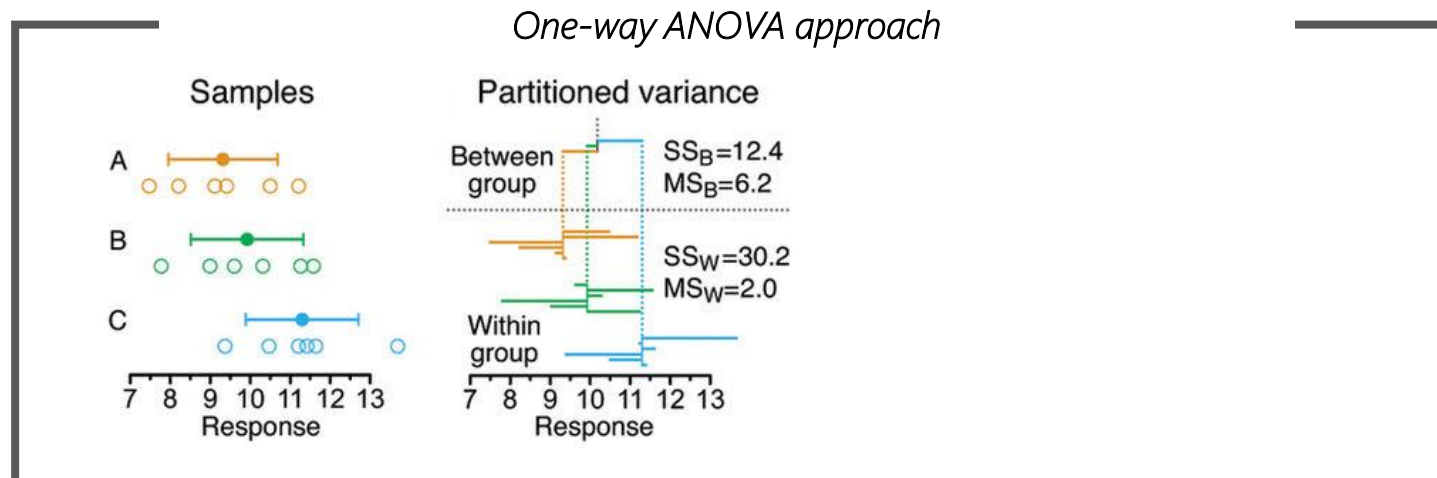
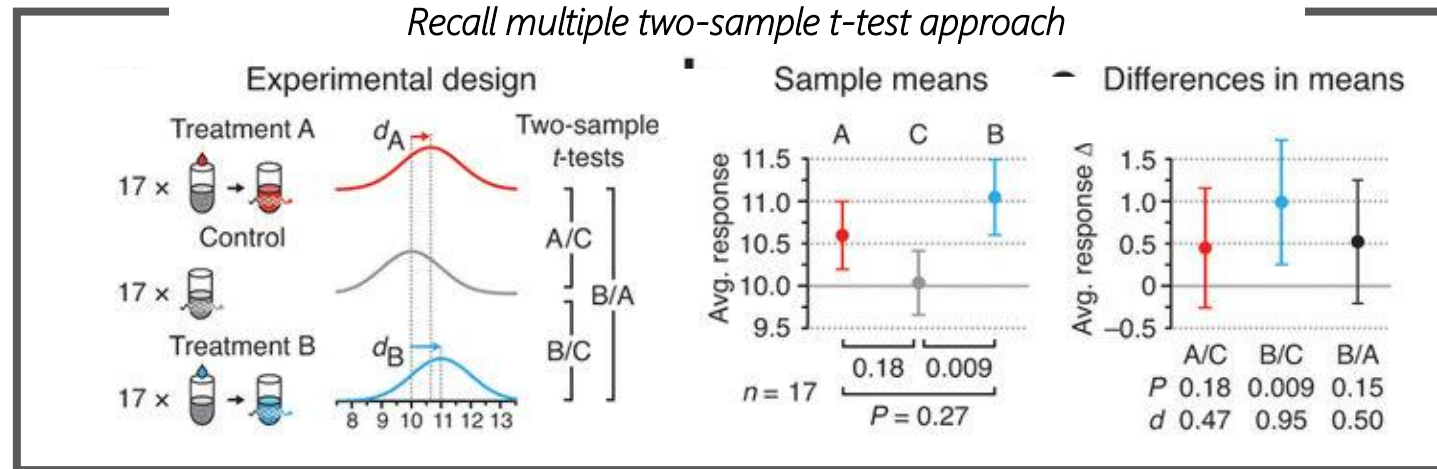
One-way ANOVA as an alternative to multiple two-sample t-tests



One-way ANOVA as an alternative to multiple two-sample t-tests

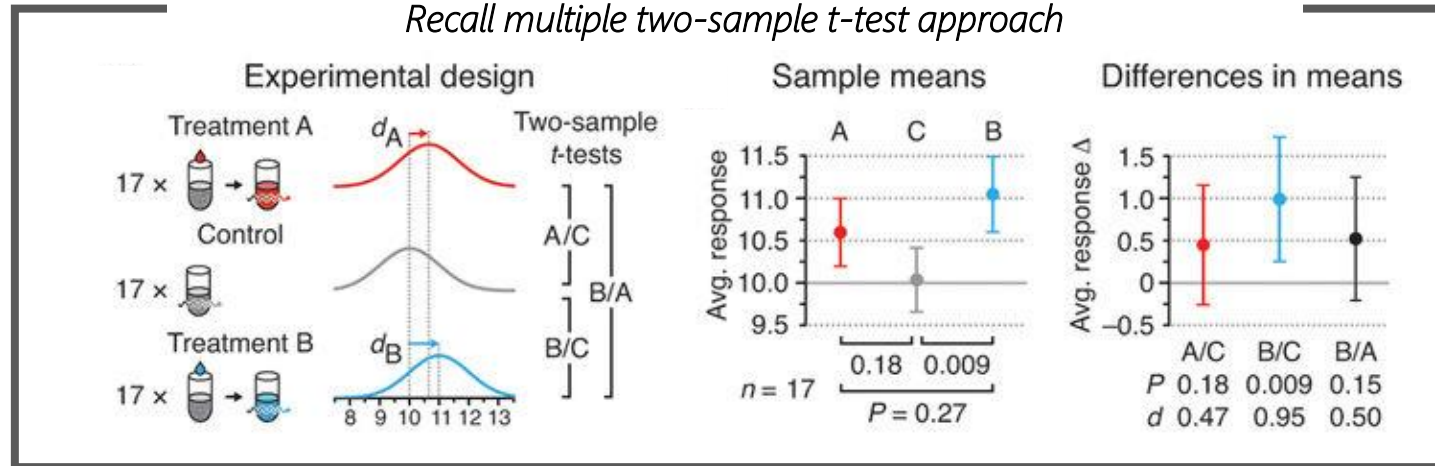


One-way ANOVA as an alternative to multiple two-sample t-tests

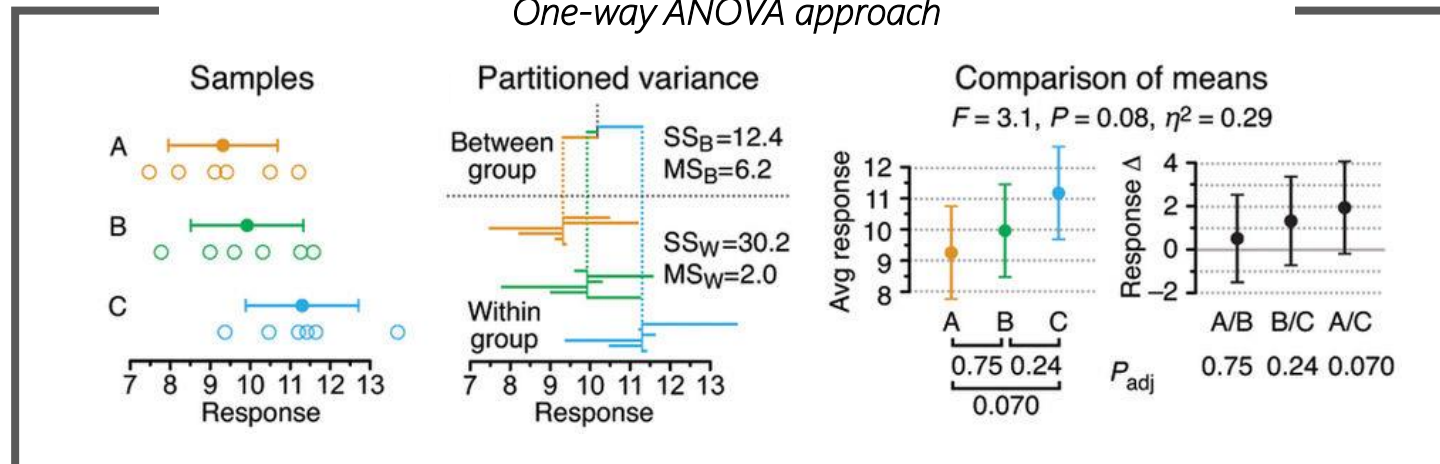


One-way ANOVA as an alternative to multiple two-sample t-tests

Recall multiple two-sample t-test approach



One-way ANOVA approach



ANOVA approaches are powerful because they include known sources of **variation**

Basic ANOVA, completely randomized design

$$\begin{aligned}
 \text{Observed feature intensity} &= \text{Systematic mean signal of disease group} + \text{Random deviation due to all sources of variation} \\
 y_{ij} &= \text{Group mean}_i + \text{Error}_{j(i)} \sim N(0, \sigma^2)
 \end{aligned}$$

Mixed effects ANOVA, with technical replicates

$$\begin{aligned}
 \text{Observed feature intensity} &= \text{Systematic mean signal of disease group} + \text{Random deviation due to individual} + \text{Random deviation due to sample preparation} + \text{Random deviation due to measurement error} \\
 y_{ijkl} &= \text{Group mean}_i + \text{Indiv}(\text{Group})_{j(i)} \sim N(0, \sigma_{\text{Indiv}}^2) + \text{Prep}(\text{Indiv})_{k(ij)} \sim N(0, \sigma_{\text{Prep}}^2) + \text{Error}_{l(ijk)} \sim N(0, \sigma_{\text{Error}}^2)
 \end{aligned}$$

Mixed effects ANOVA, with blocking

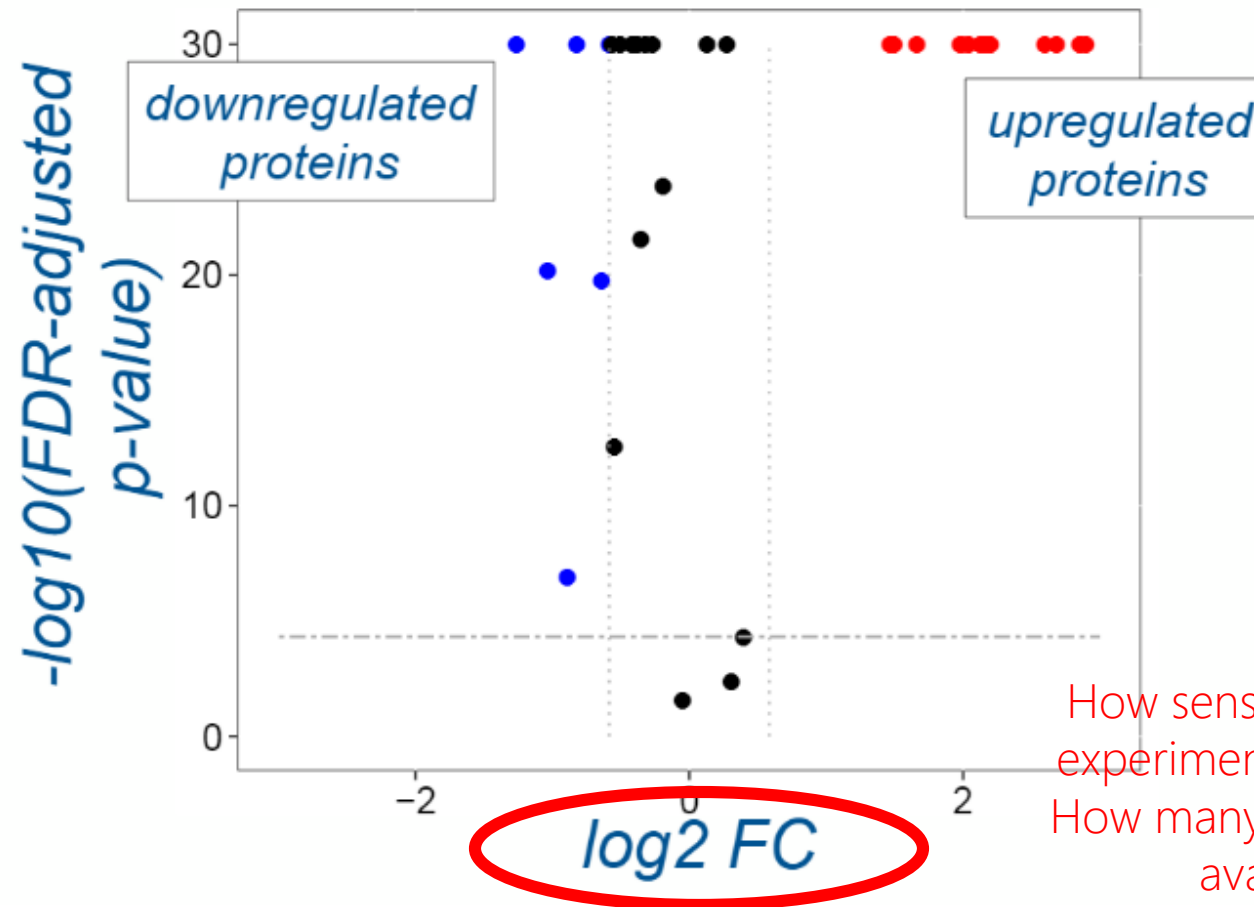
$$\begin{aligned}
 \text{Observed feature intensity} &= \text{Systematic mean signal of disease group} + \text{Random deviation due to block (e.g. plate or day)} + \text{Random deviation due to individual} + \text{Random deviation due to measurement error} \\
 y_{ijkl} &= \text{Group mean}_i + \text{Block}_k \sim N(0, \sigma_{\text{Block}}^2) + \text{Indiv}(\text{Group})_{j(i)} \sim N(0, \sigma_{\text{Indiv}}^2) + \text{Error}_{l(ijk)} \sim N(0, \sigma_{\text{Error}}^2)
 \end{aligned}$$

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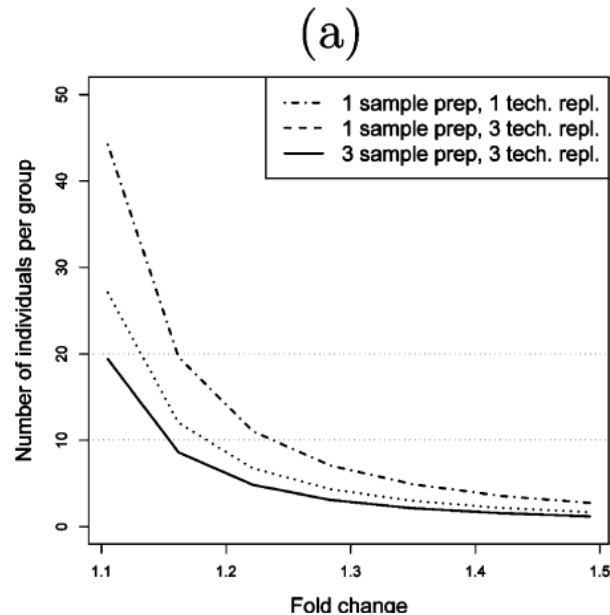
Earlier Today: Group Comparisons natively within Skyline

Statistical significance is only part of the data science story



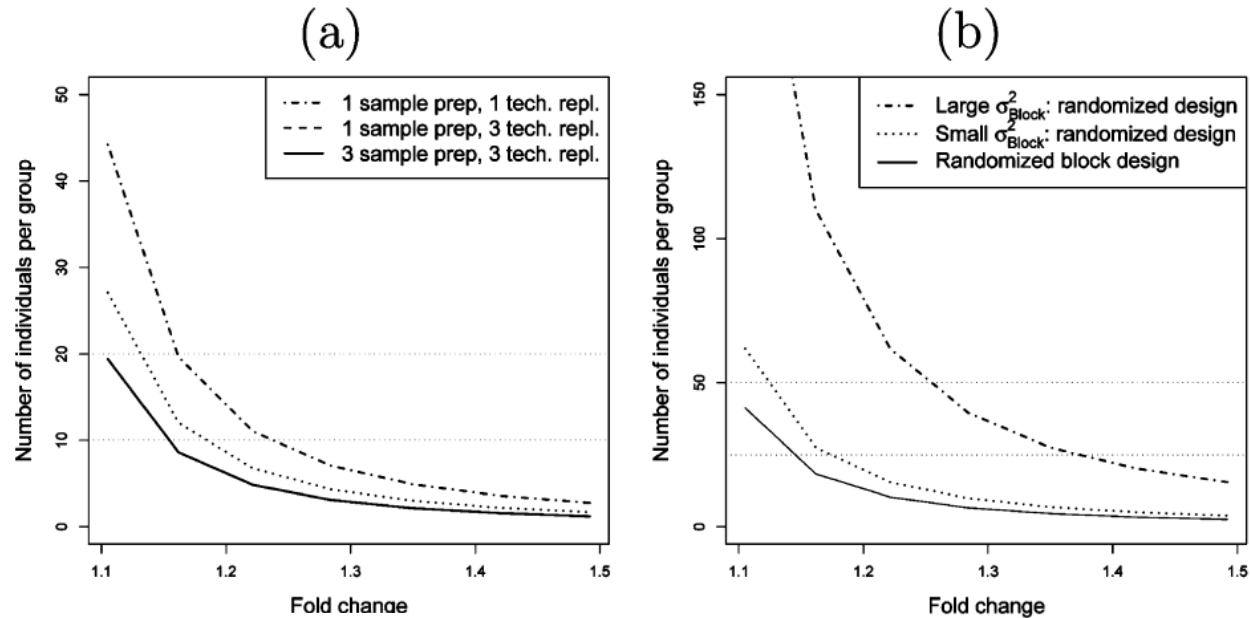
How sensitive does the experiment need to be?
How many replicates are available?

Power analysis informs how many replicates is “enough” and how sensitive the experiment is



Biological replicates are more statistically useful than technical replicates

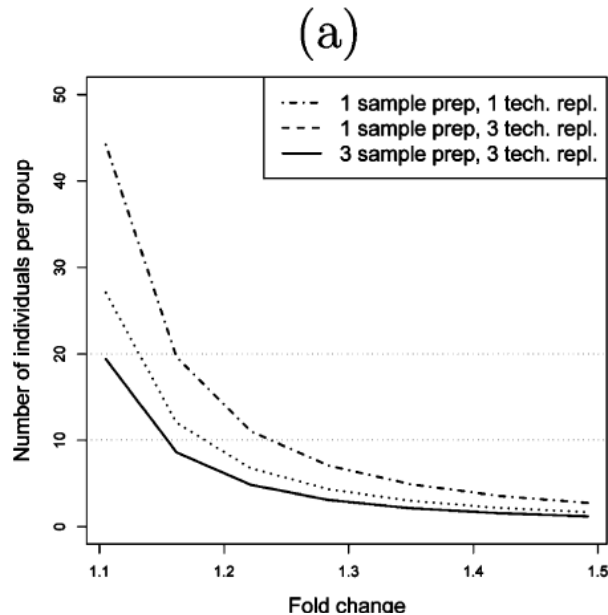
Power analysis informs how many replicates is “enough” and how sensitive the experiment is



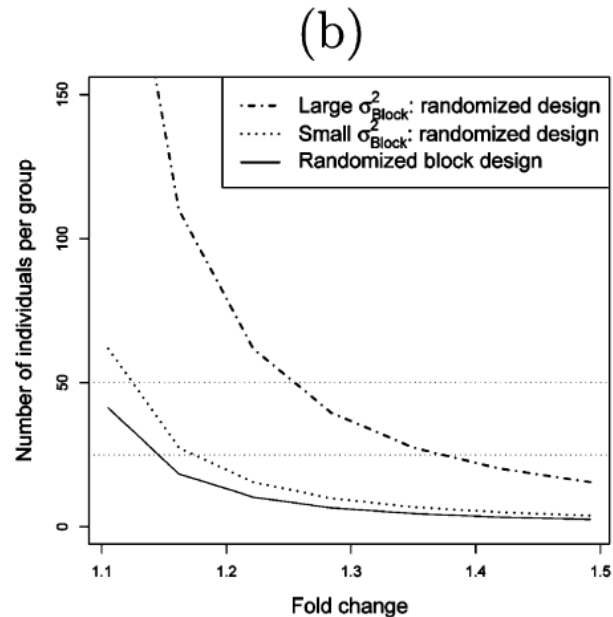
Biological replicates are more statistically useful than technical replicates

Block to minimize between-block variance

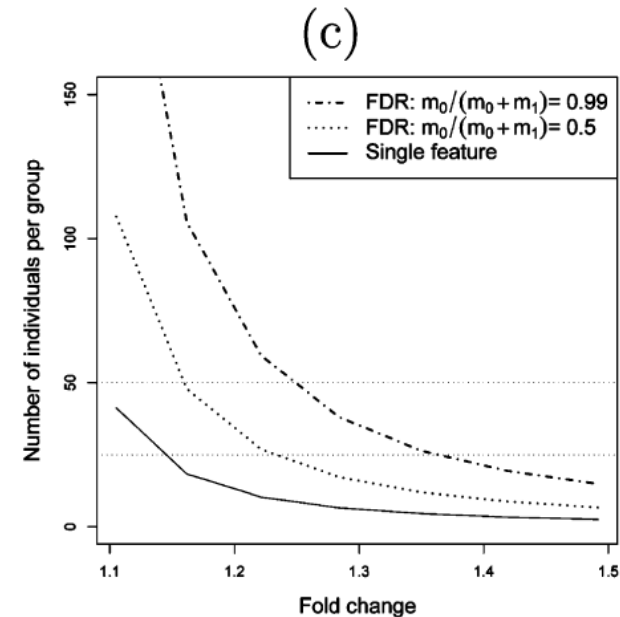
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Biological replicates are more statistically useful than technical replicates

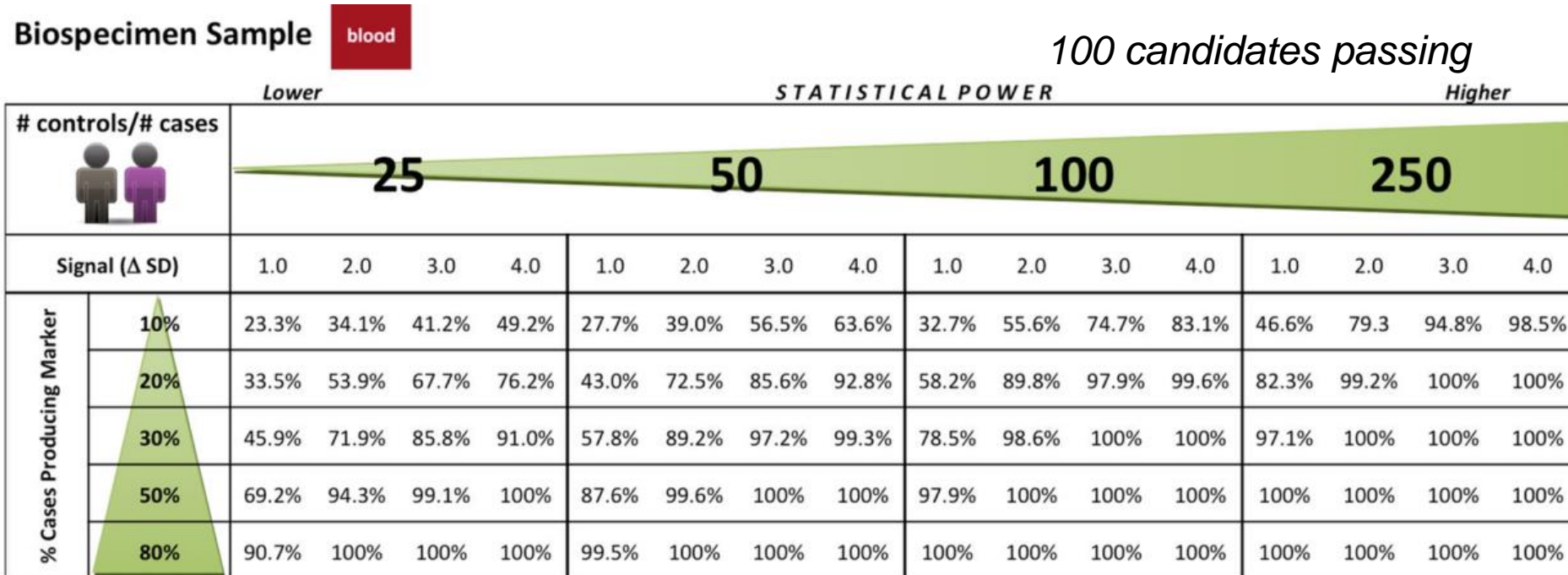


Block to minimize between-block variance

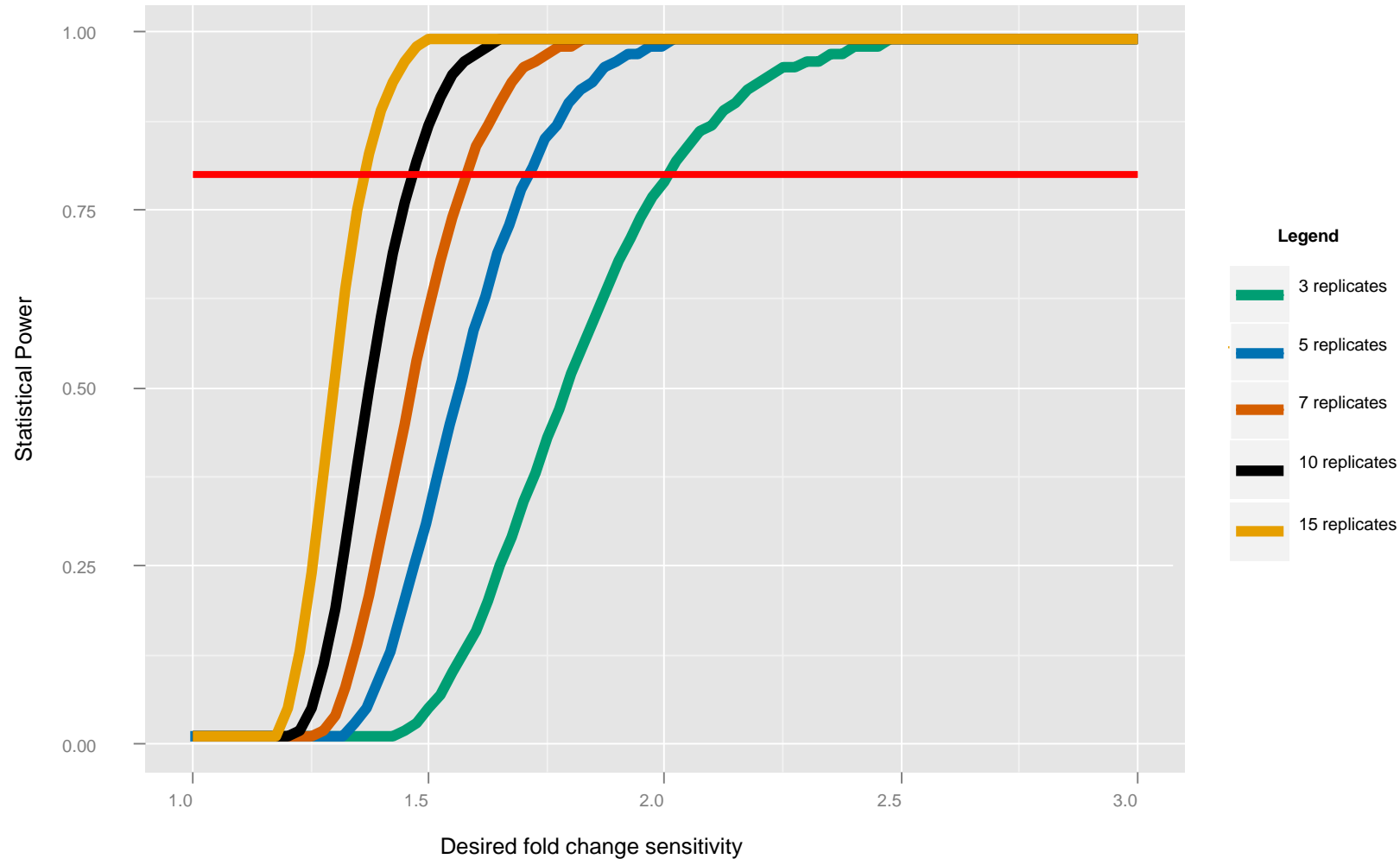


Fewer targets is better than more

Example of power analysis: biospecimen cohort size in proteomics-based biomarker discovery and verification studies



Example of power analysis: yeast proteome response to perturbation examined by DIA-MS



For more information...

Journal of
reviews **proteome**
research

Statistical Design of Quantitative Mass Spectrometry-Based Proteomic Experiments

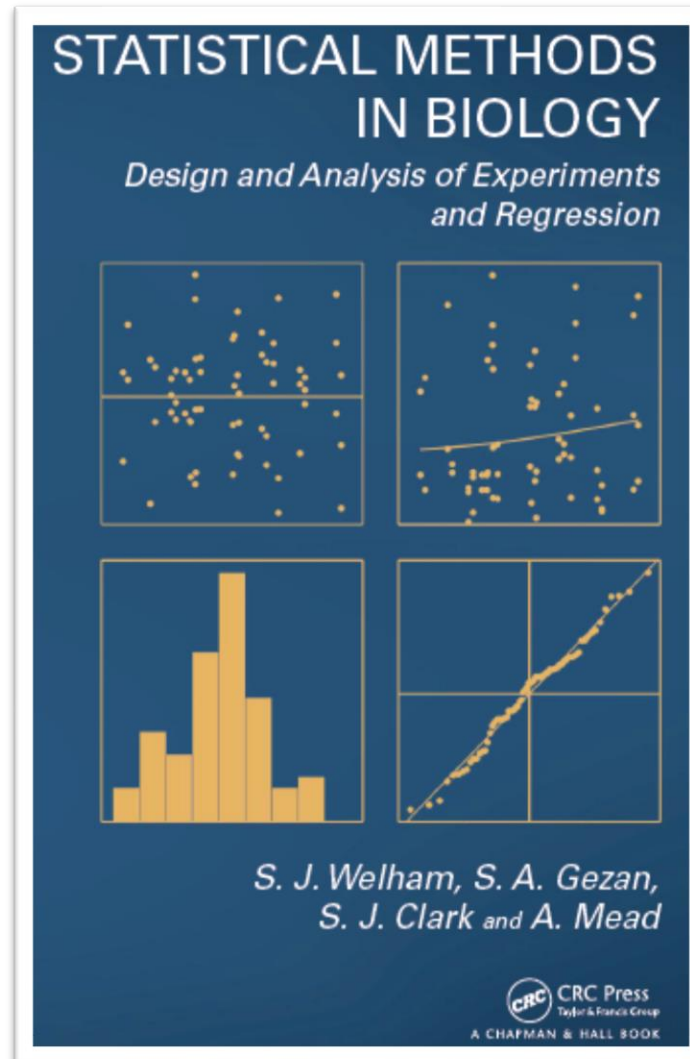
Ann L. Oberg[†] and Olga Vitek^{*,†}

Division of Biomedical Statistics and Informatics, Department of Health Sciences Research, Mayo Clinic, 200 First Street SW, Rochester, Minnesota 55905, and Department of Statistics and Department of Computer Science, Purdue University, 250 North University Street, West Lafayette, Indiana 47907

Received November 21, 2008



Since September 2013 *Nature Methods* has been publishing a monthly column on statistics called "Points of Significance." This column is intended to provide researchers in biology with a basic introduction to core statistical concepts and methods, including experimental design. Although targeted at biologists, the articles are useful guides for researchers in other disciplines as well. A continuously updated list of these articles is provided below.

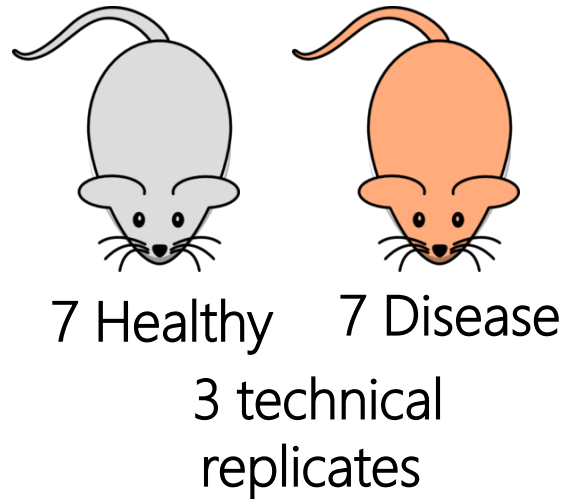


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Earlier Today: Group Comparisons natively within Skyline

Example 1: plasma proteomics in healthy vs salt-sensitive rat

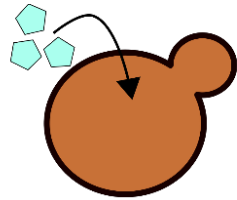


- What statistical analysis is most appropriate?
- How to block the sample preparation?

Example 2: effect of genotype + treatment on cellular aging



5 genotypes

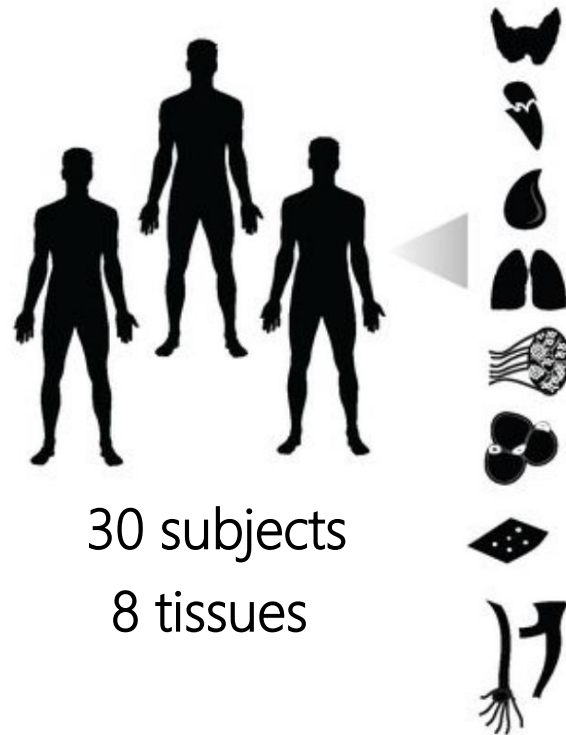


5 treatments

5 biological
replicates

- What statistical analysis is most appropriate?
- How to block the sample preparation?

Example 3: tissue-specific proteomics atlas



- What statistical analysis is most appropriate?
- How to block the sample preparation?